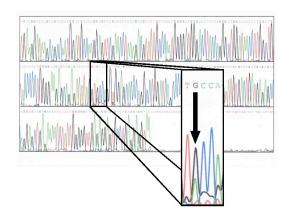
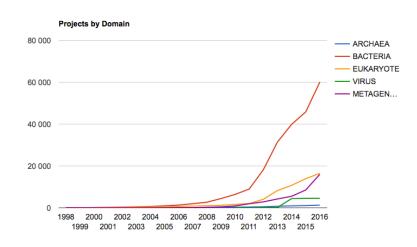
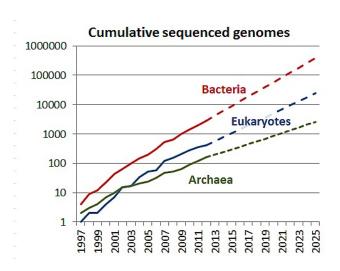
GENOME SEQUENCING

Introduction

Sequencing the genomes

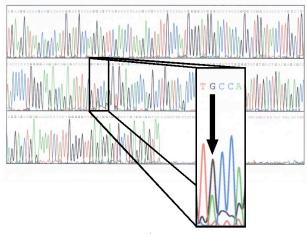


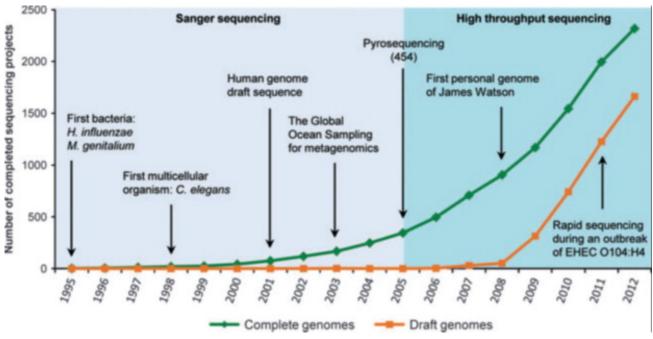




Introduction

Sequencing the genomes

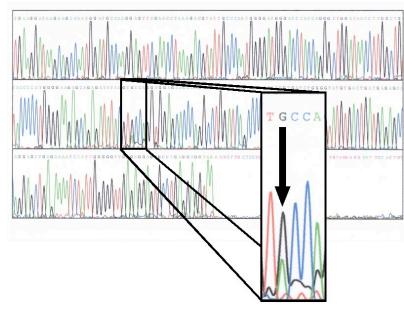




http://www.yourgenome.org/facts/timeline-organisms-that-have-had-their-genomes-sequenced ³

Introduction

Sequencing the genomes



Why sequencing these genomes?

Complete inventory of the genes of various organisms: # of genes, % of coding genome

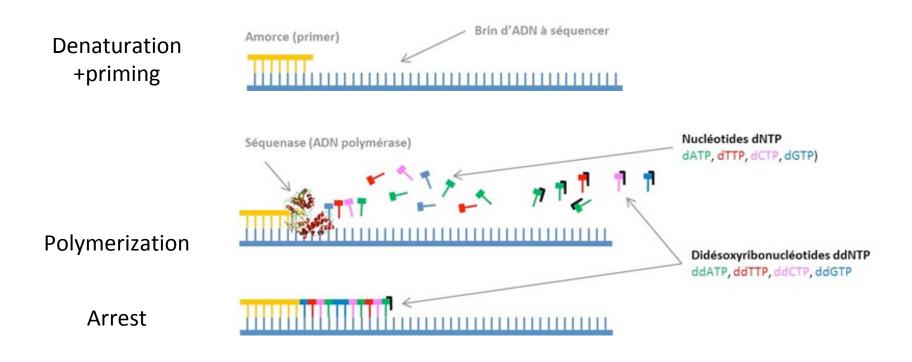
Information about the sequence of every polypeptides

Basis for new technologies and medical applications

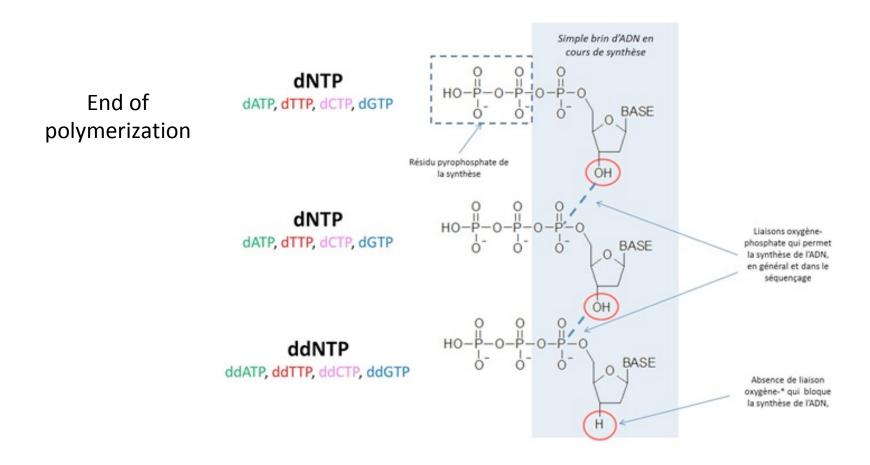
Global data at the level of the whole cell/tissue (tumor heterogeneity)

New data for evolutionnary and species classification analysis

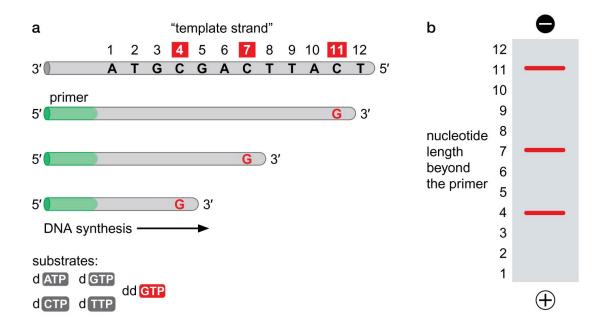
The Sanger Method (1977)

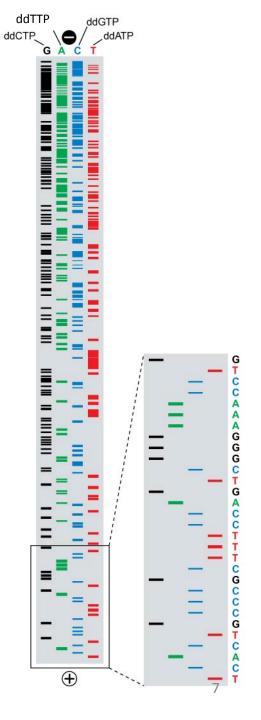


The Sanger Method (1977)

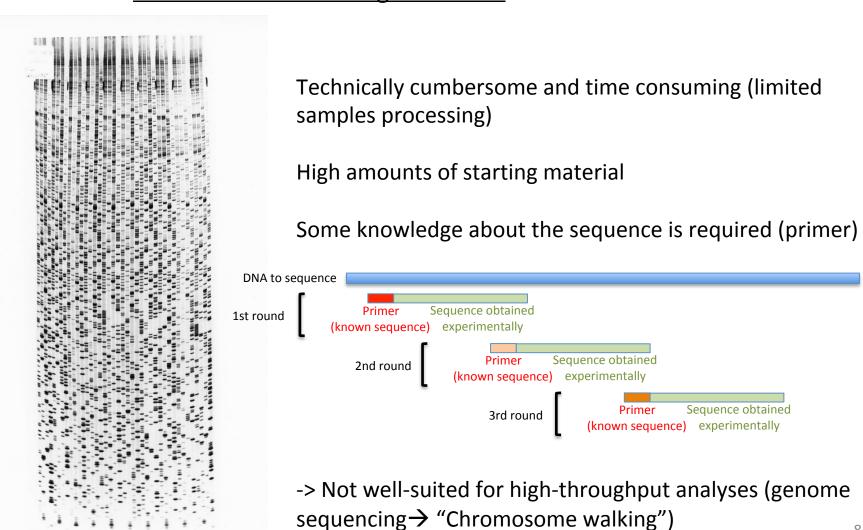


The Sanger Method (1977)



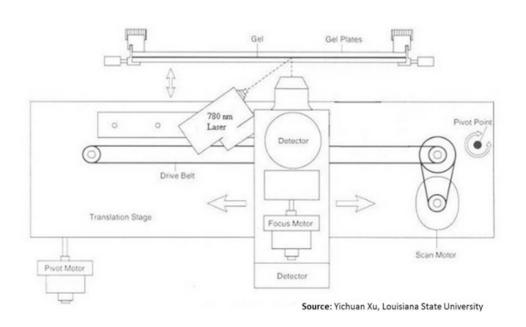


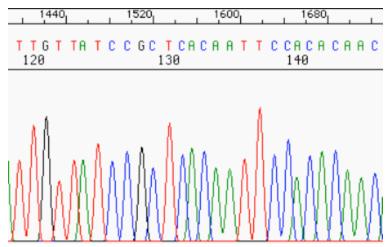
The caveats of the Sanger Method



The Sanger Method (1977)

1st generation sequencing: Improvement of sequence analysis (fluorescent ddNTPs + column electrophoresis)



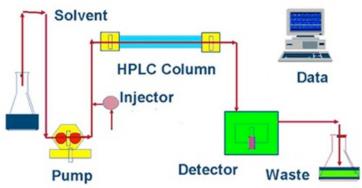


The Sanger Method (1977)

1st generation sequencing: Improvement of sequence analysis (capillary

electrophoresis)

HPLC System



Source: College of Arts and Sciences, New Mexico State University









PerkinElmer Flexar FX-15 UHPLC

Nexera UHPLC

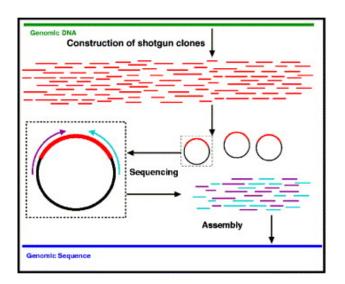
1st generation sequencing: Improvement of the methodology

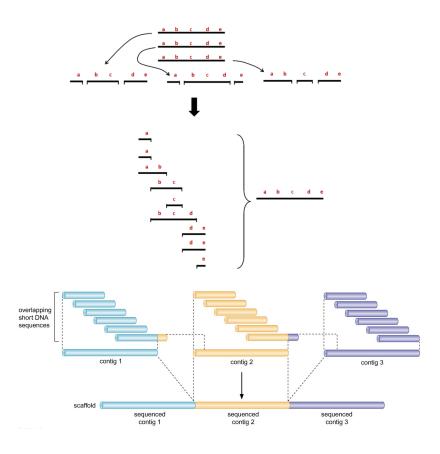
Parallel random sequencing: after fragmentation (shotgun sequencing) and cloning

- · genomic DNA is fragmented
- cloned to a plasmid vector -> transform

E. coli

 a single bacterial colony is picked -> plasmid DNA isolated





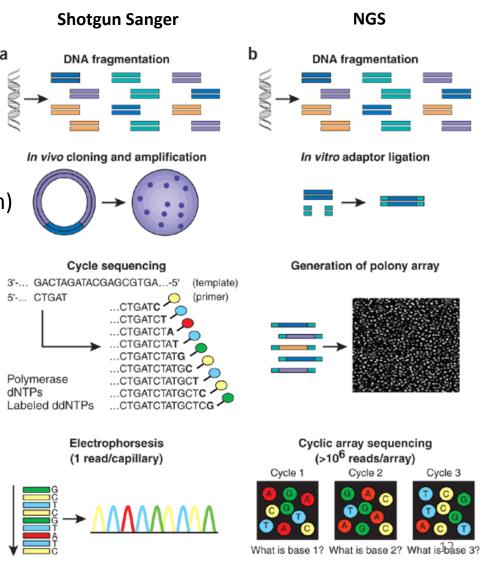
! Redundancy (8x or 20x)

2nd generation and next generation (NGS) sequencing: Areas of improvement

■ Template preparation amplification by <u>PCR</u> (Cell free system)

■ Sequencing and imaging

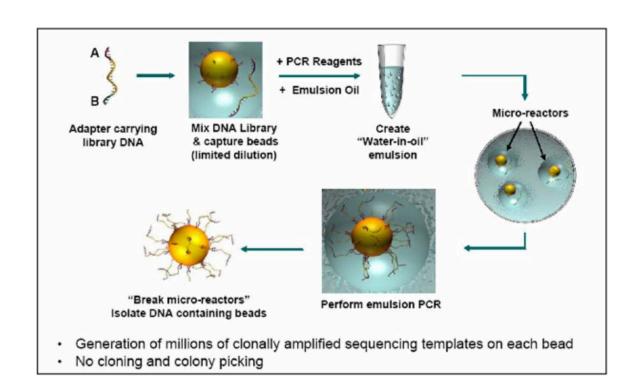
■ Data analysis



2nd generation sequencing/NGS:

Improvement of template preparation : amplification by PCR

Emulsion PCR (emPCR)



Used by 454 (life science), Polonator, Ion torrent and Solid

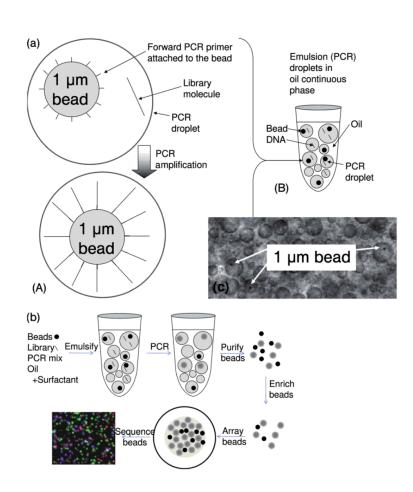
2nd generation sequencing/NGS:

Improvement of template preparation: amplification by PCR

Emulsion PCR (emPCR)

Beads are immobilized on:

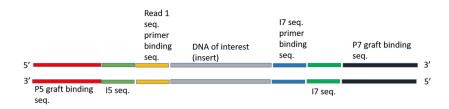
Microscope slides (Polonator)
PicoTiterPlates (PTP) (Roche/454)
Microchip sensor (Ion Torrent)

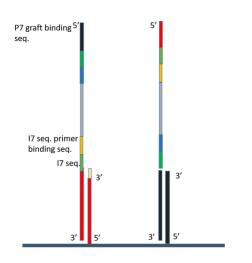


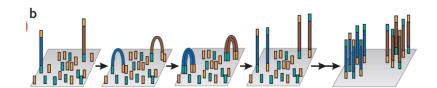
2nd generation sequencing:

Improvement of template preparation: amplification by PCR

Bridge PCR (<u>illumina plateform</u>): https://www.youtube.com/watch?v=CZeN-lgjYCo





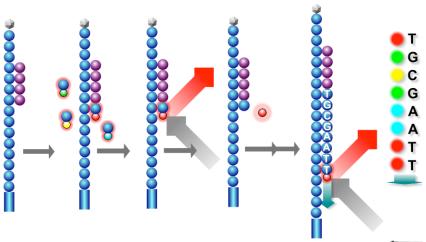


- DNA fragments are flanked with adaptors.
- A flat surface coated with two types of primers, corresponding to the adaptors.
- Amplification proceeds in cycles, with one end of each bridge tethered to the surface.
- Used by Solexa.

Fragments bound to flow cell

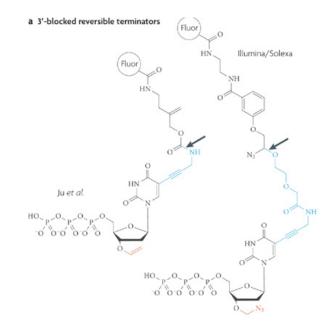
2/3rd generation sequencing : real-time sequencing

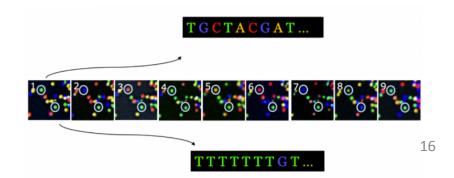
Sequencing by reversible terminator (illumina/Solexa genome Analyzer) https://www.youtube.com/watch?v=fCd6B5HRaZ8



Modified polymerase incorporates nucleotides

- after each nucleotide incorporation process stops
- camera reads fluorophore signal (filter for each nucleotide type)
- terminator and labeling is removed and cycle starts again

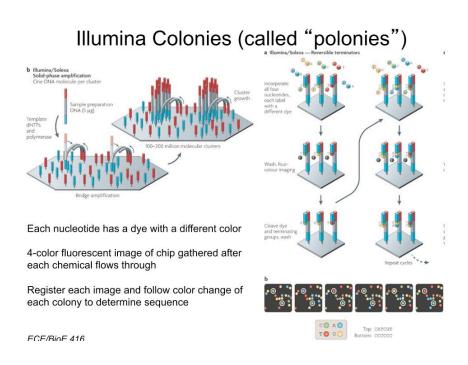




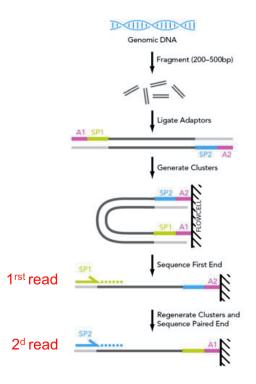
2nd generation sequencing: real-time sequencing

Sequencing by reversible terminator

Used in combination with bridge PCR in the illumina/Solexa genome Analyzer



Paired-end sequencing



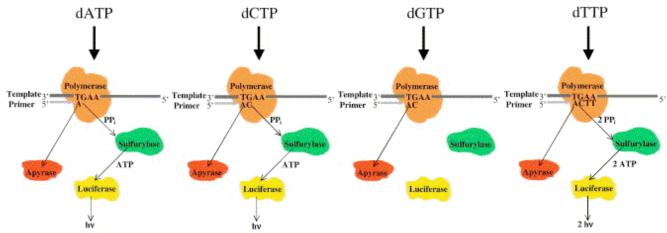
2nd generation sequencing: real-time sequencing

Sequencing by reversible terminator



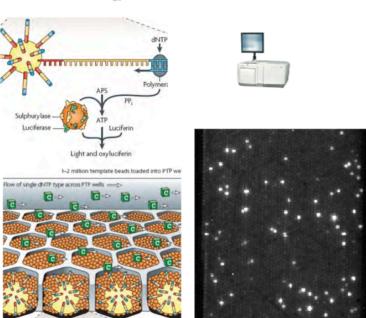
2nd generation sequencing: real-time sequencing

Pyrosequencing



Used in the 454 plateform (Roche Life Sciences)

emPCR+Pyrosequencing



2nd generation sequencing: real-time sequencing

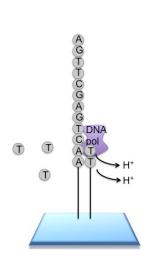


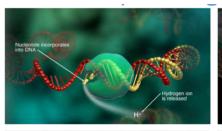


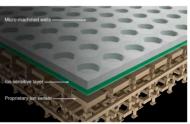
Ion torrent sequencing

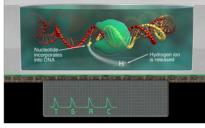
Life Technologies: Ion Proton & Ion PGM

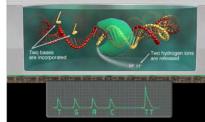
Ion proton sequencing (Ion torrent): emPCR + detection of changes in pH

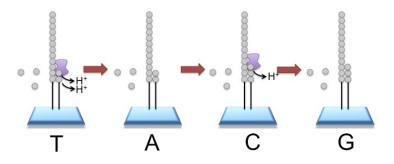


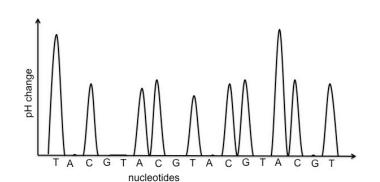












2nd generation sequencing:

Improvement of template preparation : amplification by PCR

- → Still require large amount of template
- → Sequencing of short length fragments only
- → PCR amplification induces biaises

3rd generation sequencing:

Improvement of template preparation: Single Molecule Sequencing (SMRT)

- Requires less starting material
- Provides longer sequence fragments

■ Immobilized on the solid surface by

Primers: Helicos BioSciences

Template: Helicos BioSciences

Polymerase: Pacific Biosciences, Life/Visigen, LI-

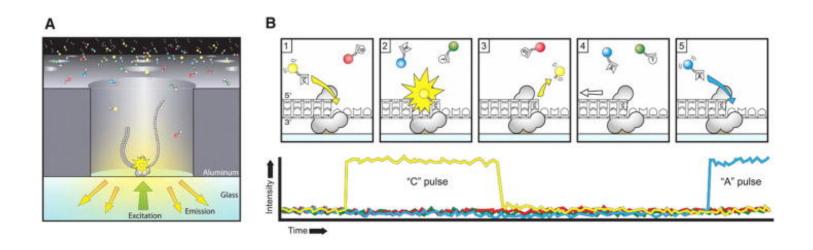
COR Biosciences

3rd generation sequencing:

Improvement of template preparation: Single Molecule Sequencing

SMRT Sequencing Technology: **Single-Molecule** Real-Time Sequencing (3rd generation)

A true continous imaging of dye-labelled nucleotides incorporation



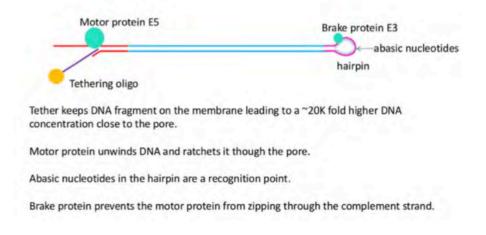
<u>Pacific Biosciences</u>: SMRT: Single Molecule Real-Time Sequencing https://www.youtube.com/watch?v=v8p4ph2MAvI

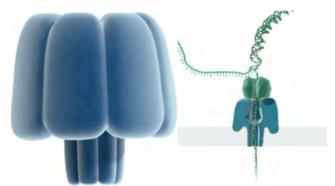
3rd generation sequencing:

Improvement of template preparation: Single Molecule Sequencing

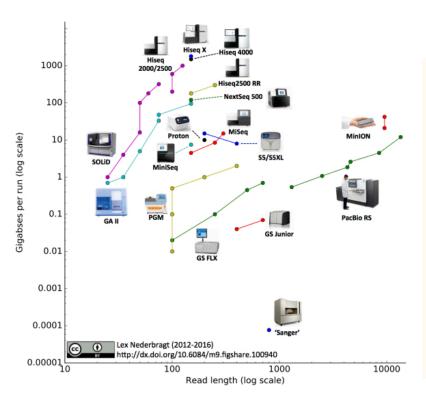
NANOPORE

Oxford Nanopore sequencing





II. Genome sequencing: the good choice



D C		c		1		
Performance	comparison	ot sec	mencing	nlattorme	Of Warious	generations
1 CHOHIHANCC	COMPanison	o_1 soc	Juchening	pianomins	or various	generations

Method	Generation	Read length (bp)	Single pass error rate (%)	No. of reads per run	Time per run	Cost per million bases (USD)	Refs.
Sanger ABI 3730×1	1st	600–1000	0.001	96	0.5–3 h	500	[14], [18], [19], [20], [21]
Ion Torrent	2nd	200	1	8.2×10^{7}	2–4 h	0.1	[15], [25]
454 (Roche) GS FLX+	2nd	700	1	1×10^6	23 h	8.57	[14], [17], [27]
Illumina HiSeq 2500 (High Output)	2nd	2 × 125	0.1	8×10^9 (paired)	7–60 h	0.03	[<u>9</u>], [<u>16</u>], [<u>26</u>]
Illumina HiSeq 2500 (Rapid Run)	2nd	2 × 250	0.1	1.2×10^9 (paired)	1– 6 days	0.04	[9], [<u>16</u>], [<u>26</u>]
SOLiD 5500×1	2nd	2 × 60	5	8×10^8	6 days	0.11	[14], [24]
PacBio RS II: P6-C4	3rd	$1.0-1.5 \times 10^4$ on average	13	$3.5-$ 7.5×10^4	0.5–4 h	0.40-0.80	[5], [12], [15]
Oxford Nanopore MinION	3rd	$2-5 \times 10^3$ on average	38	$1.1-$ 4.7×10^4	50 h	6.44–17.90	[22], [23]

II. Genome sequencing: the good choice

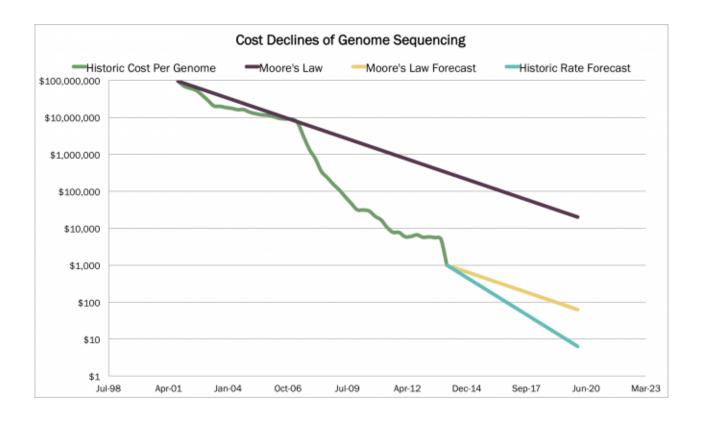
Number of sequencing machines by country

Name	Number of machines
United States	818
China	200
United Kingdom	137
Germany	135
Australia	79
Canada	74
Spain	56
Netherlands	41
France	38
Japan	34

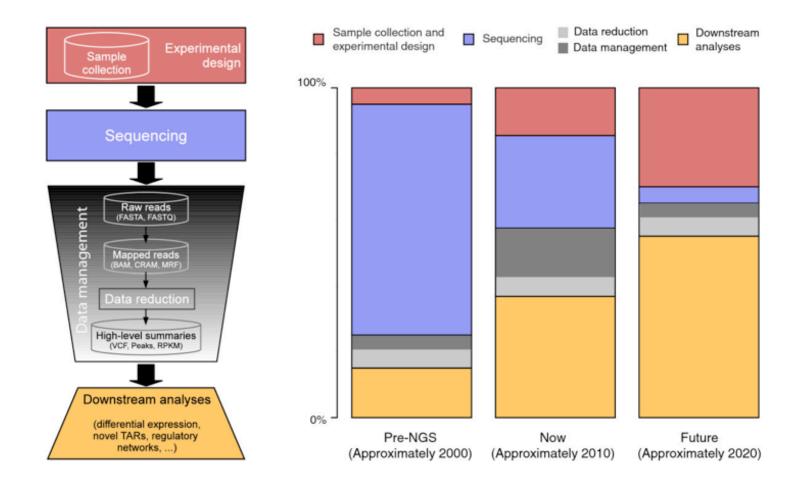
Centres with platform

Name	Number of centres
Illumina Genome Analyser 2x	279
Roche 454	265
Illumina HiSeq 2000	178
ABI SOLiD	173
Ion Torrent	101
Pacific Biosciences	26
Illumina MiSeq	23
Polonator	5

I. Genome sequencing: the price

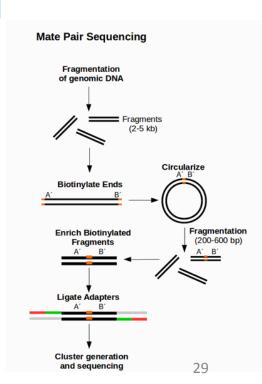


II. Data analysis



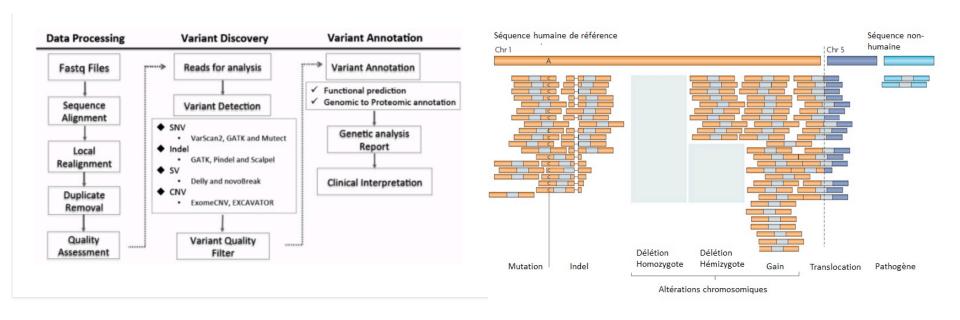
II. Data analysis

- Bioinformatics tools for:
 - Alignment
 - Base calling/polymorphism detection
 - De novo assembly
 - Genome browsing or annotation
- Challenging problems:
 - De novo assembly of short reads -> mate-paired libraries required
 - Reads in repetitive regions



III. NGS Applications

 Genome resequencing: polymorphism and mutation discovery in humans (1000
 Genomes Project)



III. NGS Applications

■ Transcriptome sequencing:

Gene expression

Alternative splicing

Transcript annotation

