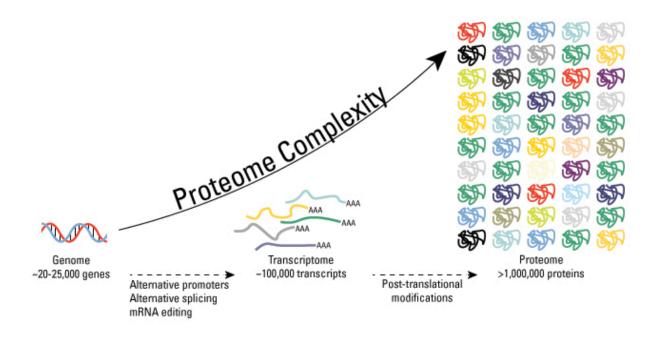


<u>Transcriptome</u>: the entire set of marked in a cell at a given time

<u>Proteome</u>: the entire set of polypeptides found in a cell at a given time

<u>Interactome</u>: the set of protein-protein interactions in a cell at a given time



II. Principes de la régulation de l'expression des gènes

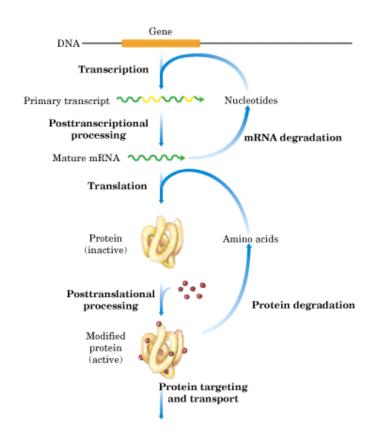
Principle #1: constitutive vs regulated gene expression

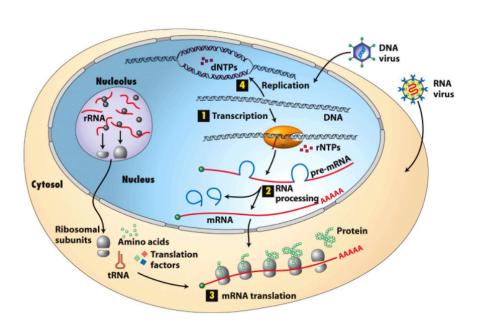
There are approx 10.000 genes expressed in an eucaryotic cell (4.000 in yeast)

Most of the genes code for polypeptides that are required at all times at a more or less constant level, expressed at low levels, common to different cell types (housekeeping genes)

the minority of mARNa are expressed at high levels, they are specific to the cell type or conditions (*luxury genes*)

Principle #2: Gene regulation can be acheived at several levels along polypeptide synthesis (different in prokaryotes and eukaryotes).





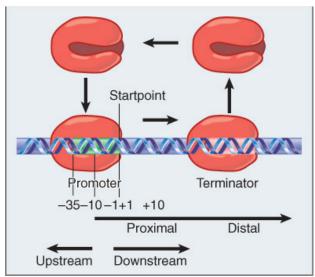
Principle #3: Transcription proceeds through a series of defined processes: inititation, elongation and termination

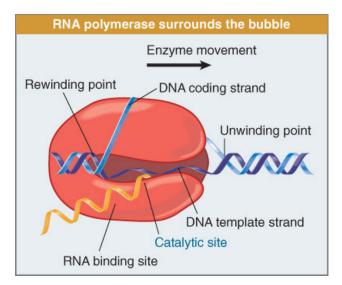
RNA polymerase recognizes **promoter** sequences (initiation).

RNA polymerase synthetizes RNA complementary to the template DNA (elongation)

RNA synthesis proceeds through terminator sequences (**termination**)

The promoter, the transcribed region and the terminator define a **transcription unit**



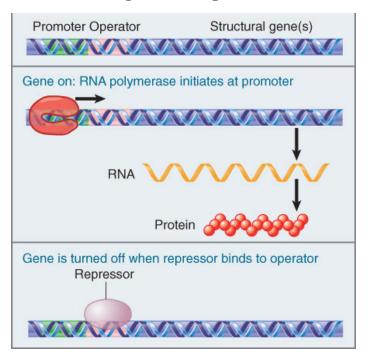


Principe #4: Transcription initiation is regulated **by trans-acting proteins** (activators or repressors) that bind **to cis-acting sites** in or near promoters

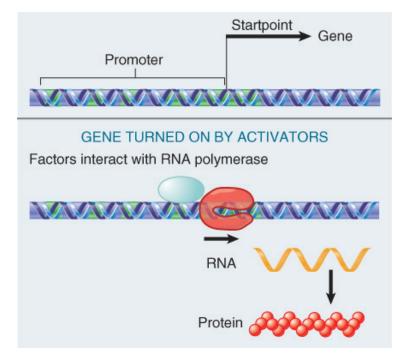
Repressors bind to operators in DNA to prevent or reduces the RNA polymerase activity: (i.e., negative regulation)

Activator binds to DNA sites called enhancer to enhance the RNA polymerase activity. (i.e., positive regulation)

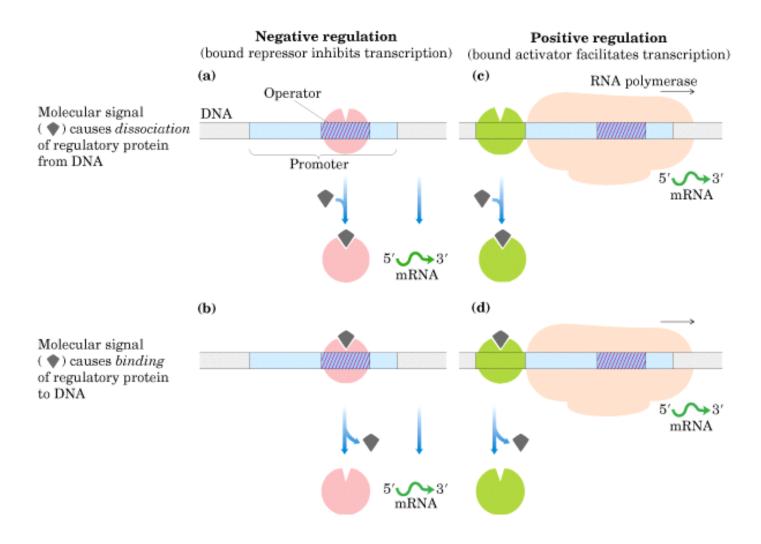
Negative regulation



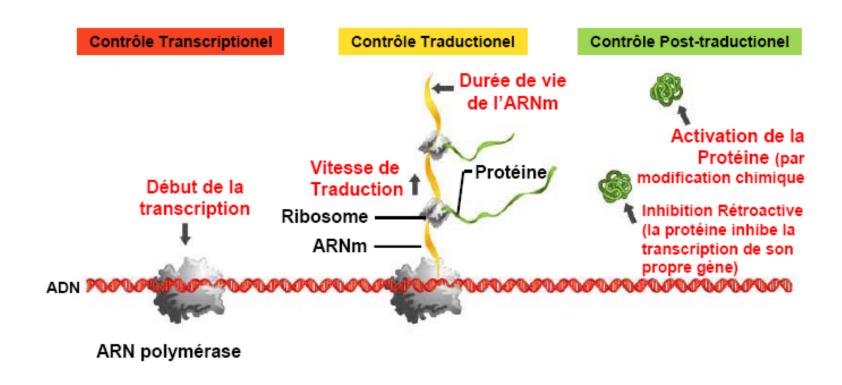
Positive regulation



Principle #5: Activation or repression by activators or repressors can be regulated by **co-regulators** (small molecules in procaryotes)



Principle: Gene regulation in Prokaryotes is achieved at the **transcription initiation** level



Prokaryotic genes are expressed and regulated as operons



Operator: DNA sequences (binding sites) involved in transcription initiation control

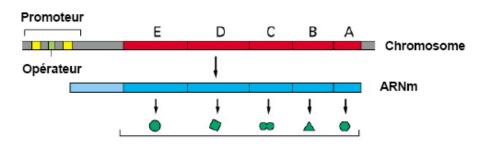
Promoter: binding of RNA polymerase

+1: transcription start

RBS: ribosome binding site

CDS: coding sequence

Terminator: ends transcription



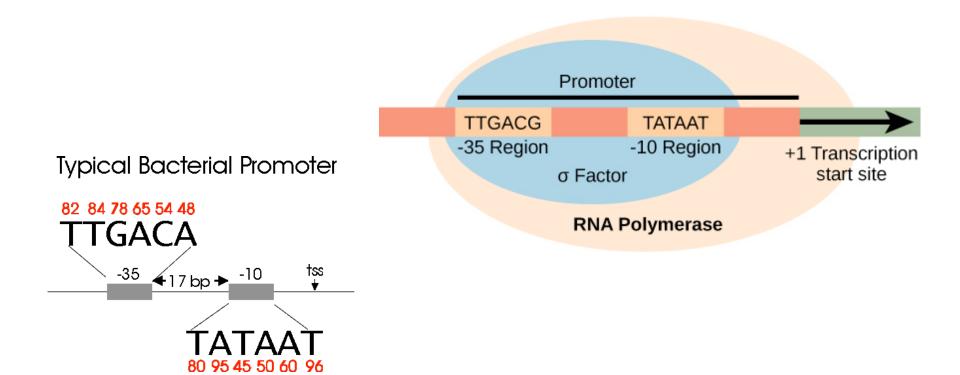
Bacterial promoters are defined by 4 core elements

TSS: transcription start site (purine in >90%)

TATA box at around -10 from the TSS

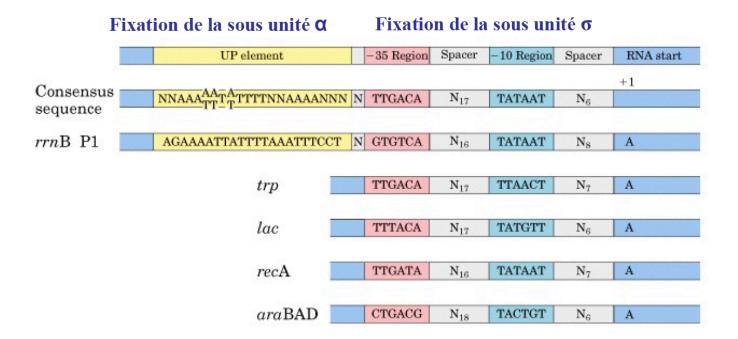
-35 Box

An insert region between the boxes of 16-18 nucleotide



Divergence from canonical sequences and structure defines how efficiently a promoter will initiate transcription (promoter strength : weak or strong).

Mutations in promoter regions can affect promoter strength (mutations up/down)



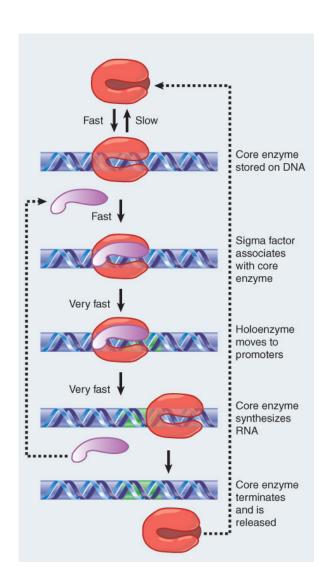
RNA polymerase requires σ -factors for efficient binding to DNA-sequences at TSS

The σ -factor has specific domains that recognize canonical elements in the promoter

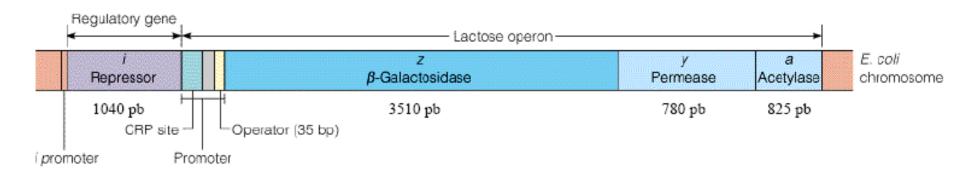
The σ -factor controls the affinity of the polymerase for the promoter sequences

Recruitment of RNA polymerases to TSS is regulated by activating and repressing trans-acting factors

Most cis-acting elements are close to the TSS



Lactose operon encodes enzymes involved in β -galactosides catabolism



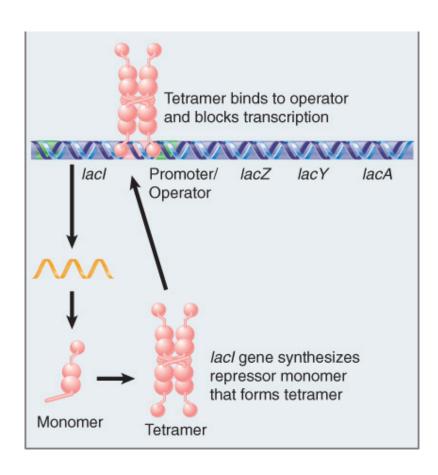
LacZ : β -galactosidase (hydrolyzes β -galactoside)

LacY : Permease (imports β -galactosides inside *E. coli*)

LacA : trans-Acetylase (acetylates β -galactosides)

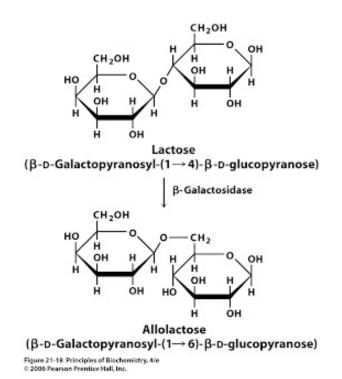
The *lac repressor* is transcribed from its own promoter Pi

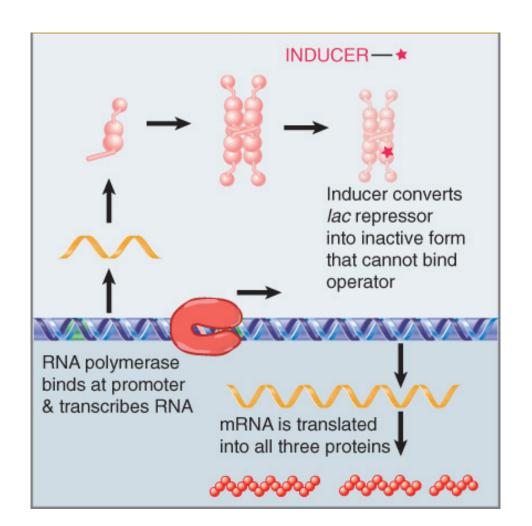
Lacl binds the Plac (operator) and prevents transcription initiation



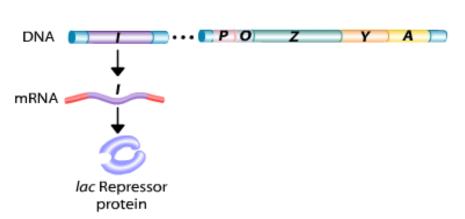
Lacl is itself inactivated by an inducer (allolactose)

Lactose paradox

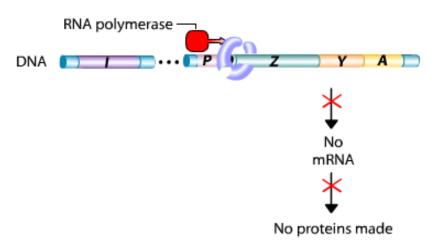




Glucose/ No Lactose



The Lac repressor protein, encoded by the *I* gene, is expressed in the absence or presence of lactose.



In the absence of lactose, the Lac repressor binds to the *lac* operator site.

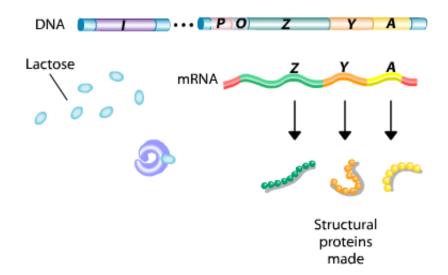
Since RNA polymerase is unable to transcribe the *lac* structural genes, the corresponding proteins are not made.

<u>Lactose</u>



When lactose is present in the cell medium, it binds to the allosteric site of the Lac repressor. This changes the conformation of the repressor.

In this conformation, the repressor can no longer bind to the *lac* operator site.

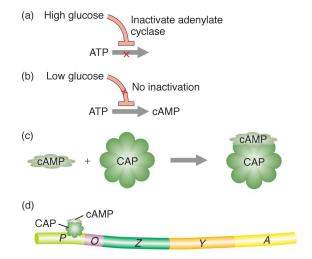


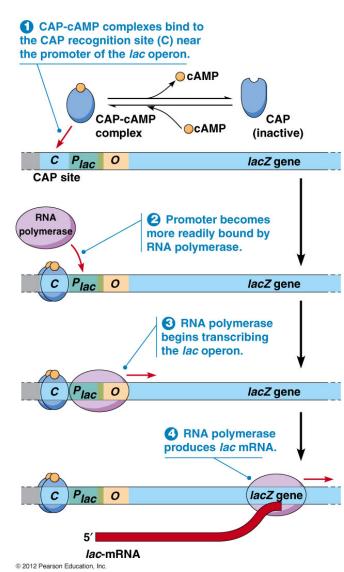
Without the repressor blocking its way, RNA polymerase is able to transcribe the structural genes.

Thus, in the presence of lactose, the *lac* structural genes are expressed. The proteins encoded by the *Z* and *Y* genes are required for the metabolism of lactose.

Preferential use of Glucose over lactose is acheived through the **cAMP Receptor Complex**

(CRP)



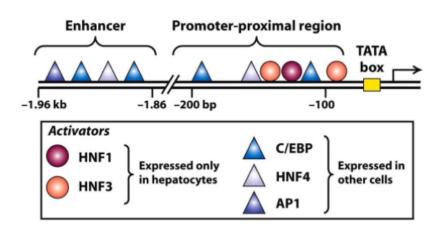


Unlike bacterial cells and most single cell eukaryotes, cells in multicellular organisms have relatively few genes that are directly and reversibly regulated by environmental conditions

Instead multicellular organisms have many different, specialized cells. Hence, **tissue-specific gene control** is important for development and differentiation.

every gene has more than one gene regulator (all of which must be on for the gene to function).

Eukaryotic regulatory elements are usually several Kb away from the promoter



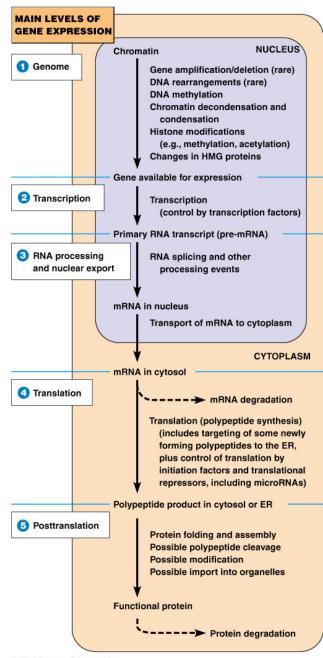
Genome Level (Chromatin Remodelling and DNA rearrangements)

Transcriptional Control

Post-Transcriptional Control

Translational Control

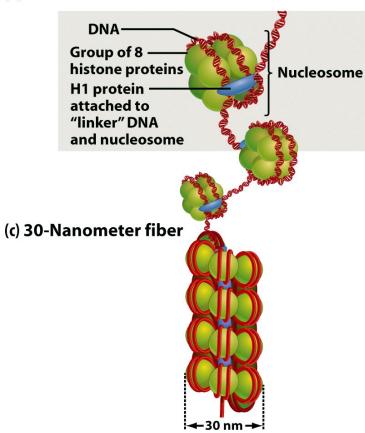
Post-Tranlational Control (protein activity control)



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III.1 Chromatin structure

(b) Nucleosome structure

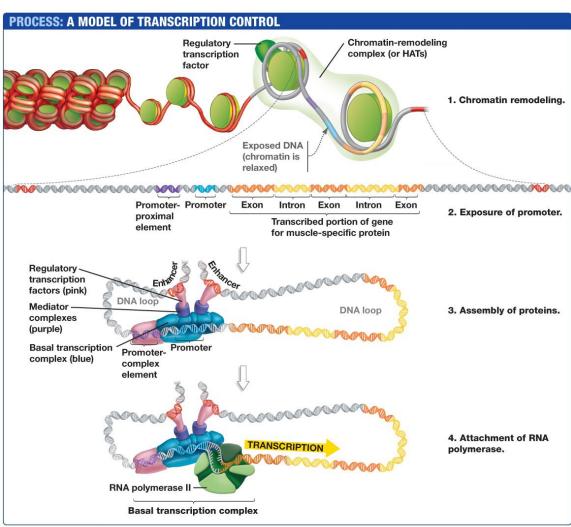


III.2 Transcriptional control

Three Classes of Proteins Are Involved in Transcriptional Activation

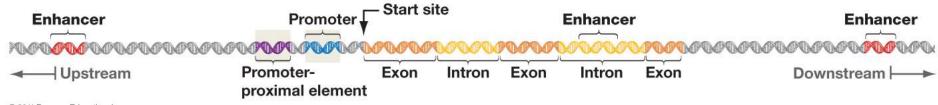
basal transcription factors (TBP)

DNA-binding transactivators, Coactivators (Mediator).



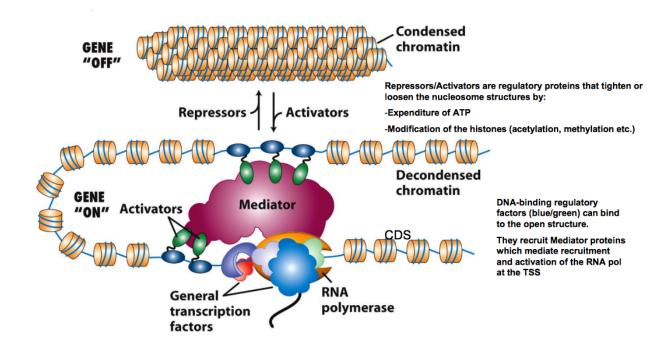
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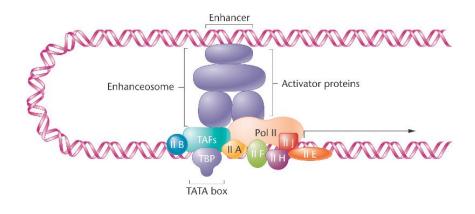
Eukaryotic regulatory sequences can be close or far from the core promoter



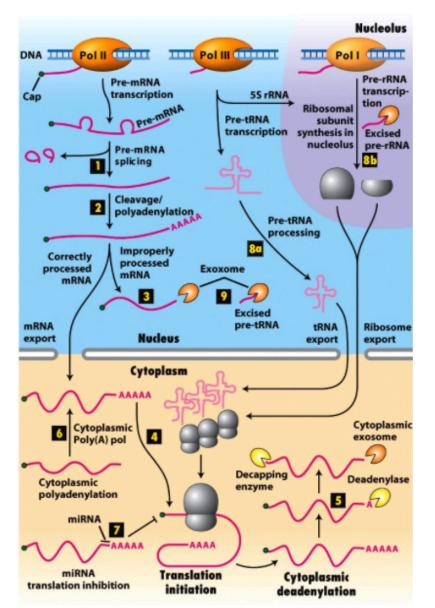
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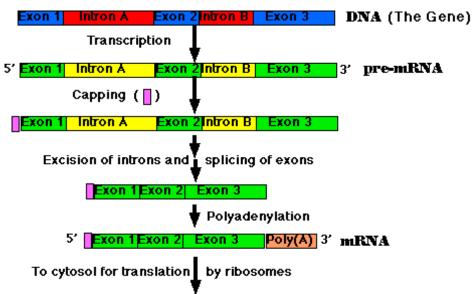
Transcription factors are key regulators of eukaryotic gene expression





III.3 Post-transcriptional control of gene expression





III.3 Post-transcriptional control of gene expression: non coding RNA

only 1% of the genome is translated into proteins but much more transcribed

A great portion of the (**procaryotic and eukaryotic**) genome is transcribed into non coding RNAs. Some of which can be involved in gene regulation :

- sncRNA
- LncRNA
- miRNAs

SUMMARY TABLE 18.1 Regulating Gene Expression in Bacteria and Eukaryotes

Level of Regulation	Bacteria	Eukaryotes		
Chromatin remodeling	 Limited packaging of DNA Remodeling not a major issue in regulating gene expression. 	 Extensive packaging of DNA Chromatin must be opened for transcription to begin. 		
Transcription	 Positive and negative control by regulatory proteins that act at sites close to the promoter Sigma interacts with promoter. 	 Positive and negative control by regulatory proteins that act at sites close to and far from promoter Large basal transcription complex interacts with promoter. Mediator complex required. 		
RNA processing	None documented	 Extensive processing: alternative splicing of introns addition of 5' cap and 3' tail 		
mRNA stability	Some RNA interference documented	 For many genes, RNA interference limits life span or translation rate. 		
Translation	 Regulatory proteins bind to mRNAs and/or ribosome and affect translation rate. 	 Regulatory proteins bind to mRNAs and/or ribosome and affect translation rate. 		
Post-translational modification	 Folding by chaperone proteins Chemical modification (e.g., phosphorylation) may change activity. 	 Folding by chaperone proteins Chemical modification (glycosylation, phosphorylation) Ubiquination targets proteins for destruction by proteasome. 		

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IV.1 SAGE Technology

SAGE: Serial *analysis of gene expression (1995)*:

Unic tags are used to identify each mRNA

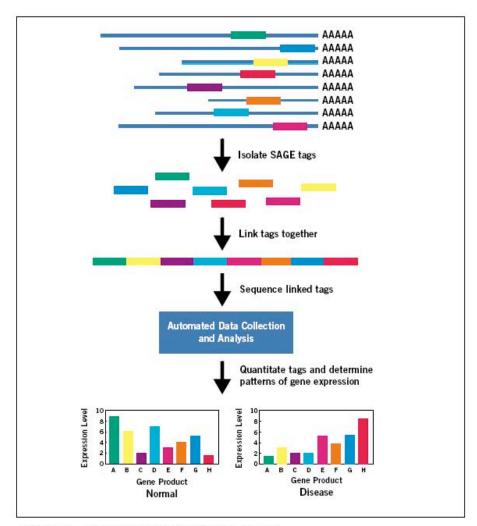
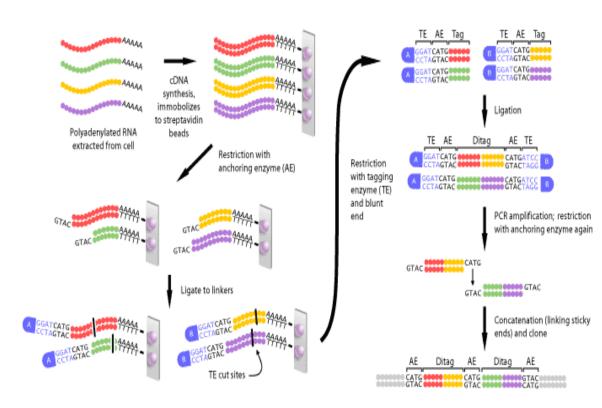


Figure 1. Schematic of SAGE method (Courtesy of sagenet.org)3

IV.1 SAGE Technology

- 1. Isolate the mRNA of an input sample (e.g. a tumour).
- 2. Extract a small chunk of sequence from a defined position of each mRNA molecule.
- Link these small pieces of sequence together to form a long chain (or concatamer).
- 4. Clone these chains into a **vector** which can be taken up by bacteria.
- 5. Sequence these chains using modern high-throughput DNA sequencers.
- 6. Process this data with a computer to count the small sequence tags.



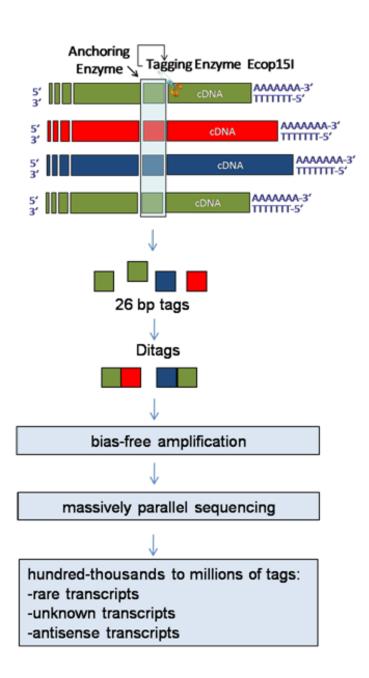
IV.1 SAGE Technology

TAG	COUNT	TAG	COUNT	TAG	COUNT
CCCATCGTCC	1286	CACTACTCAC	245	TTCACTGTGA	150
CCTCCAGCTA	715	ACTAACACCC	229	ACGCAGGGAG	142
CTAAGACTTC	559	AGCCCTACAA	222	TGCTCCTACC	140
GCCCAGGTCA	519	ACTTTTTCAA	217	CAAACCATCC	140
CACCTAATTG	469	GCCGGGTGGG	207	CCCCCTGGAT	136
CCTGTAATCC	448	GACATCAAGT	198	ATTGGAGTGC	136
TTCATACACC	400	ATCGTGGCGG	193	GCAGGGCCTC	128
ACATTGGGTG	377	GACCCAAGAT	190	CCGCTGCACT	127
GTGAAACCCC	359	GTGAAACCCT	188	GGAAAACAGA	119
CCACTGCACT	359	CTGGCCCTCG	186	TCACCGGTCA	118
TGATTTCACT	358	GCTTTATTTG	185	GTGCACTGAG	118
ACCCTTGGCC	344	CTAGCCTCAC	172	CCTCAGGATA	114
ATTTGAGAAG	320	GCGAAACCCT	167	CTCATAAGGA	113
GTGACCACGG	294	AAAACATTCT	161	ATCATGGGGA	110

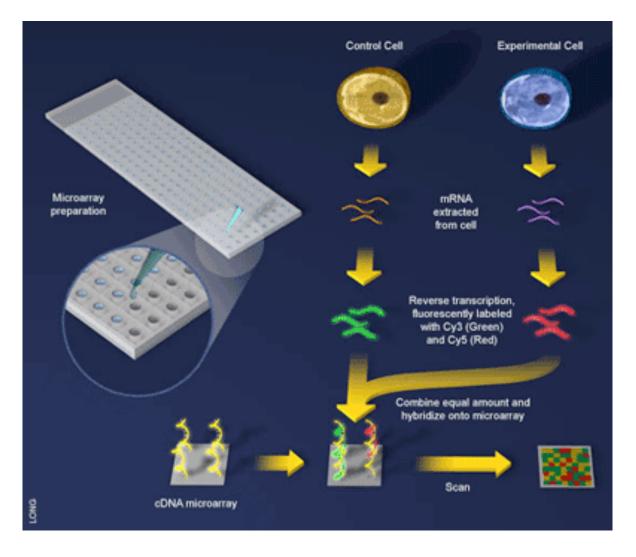
- Collect sequence records from GenBank
- Assign sequence orientation (by finding poly-A tail or poly-A signal or from annotations)
- Assign UniGene identifier to each sequence with a SAGE tag
- Record (for each tag-gene pair)
 - #sequences with this tag
 - #sequences in gene cluster with this tag
- Softwares : <u>SAGE Genie</u> or SAGE map

IV.1 SAGE Technology

HT-SuperSAGE



IV.2 Microarray Technology

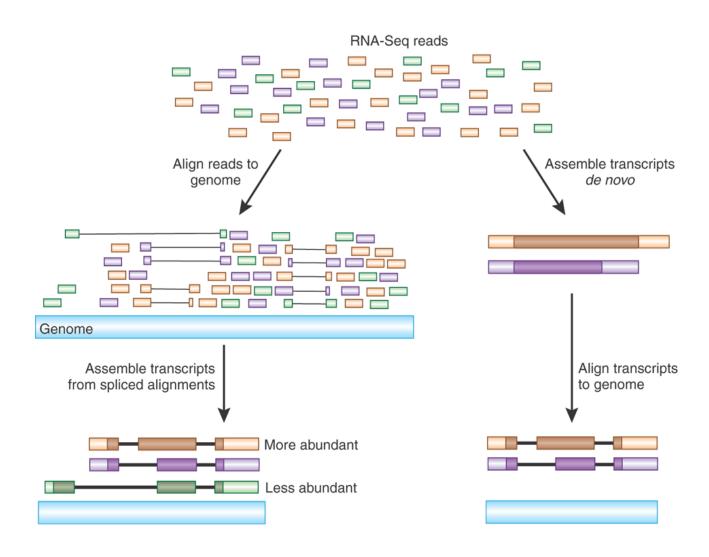


Microarray technology animation

IV.2 Microarray Technology

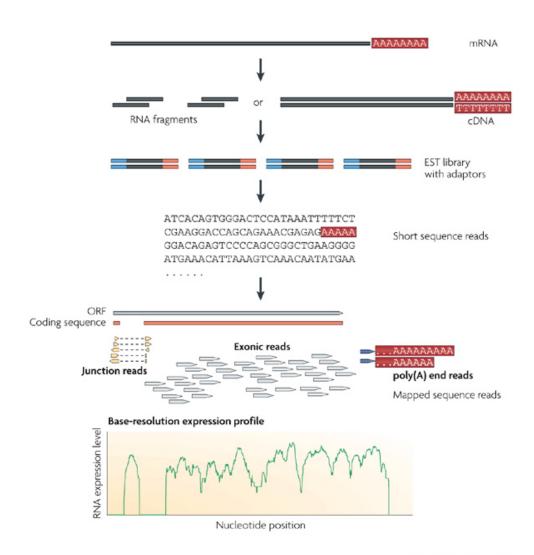
Features	SAGE	Microarray	
Detects unknown transcripts	Yes	No	
Quantification	Absolute measure	Relative measure	
Sensitivity	High	Moderate	
Specificity	Moderate	High	
Reproducibility	Good for higher abundance transcripts	Good for data from intra-platform comparison	
Direct cost	5-10X higher than arrays.	5-10 X lower than SAGE	

IV.3 RNA-Sequencing (RNA-Seq) : basics



IV.3 RNA-Sequencing (RNA-Seq): methodology

- 1. RNA extraction
- 2. Reverse transcription
- 3. Library preparation
- 4. Polymerase Chain reaction
- 5. Sequencing
- 6. Bioinformatic processing



IV.3 RNA-Sequencing (RNA-Seq)

Detection of novel transcripts and other changes

Dynamic range

Specificity and sensitivity

Costly and requires high computational power