

# **Bioinformatics softwares (R/PLINK)**

**GBIO0002**

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# Outline

- **R**
- **Features of R**
- **Simple Data Visualization**
- **Advance Data Visualization**
- **Data Transformation**
- **PLINK**

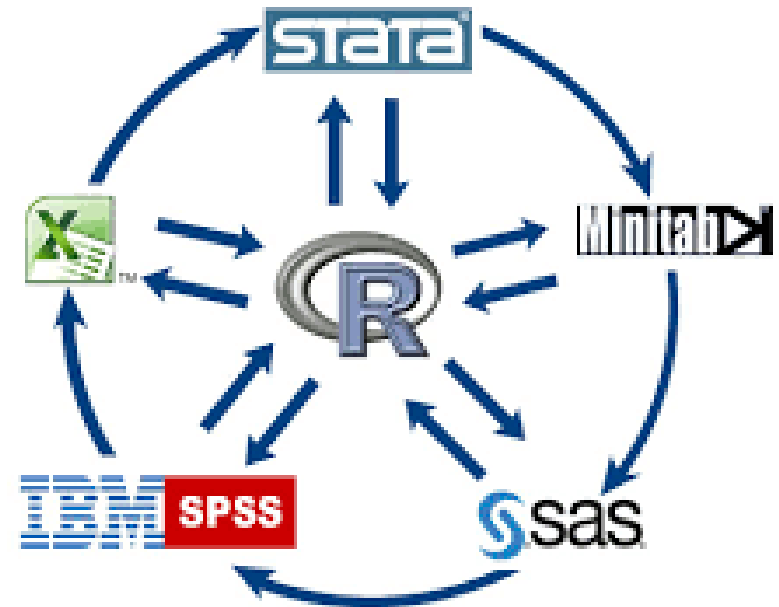
Introduction to



**A basic tutorial**

# Definition of R

- R is a free software environment for statistical computing and graphics
- R is considered to be one of the most widely used languages amongst statisticians , data miners, bioinformaticians and others.
- Other commercial statistical packages are SPASS , SAS, Matlab



# Statistical language GUI

The SAS GUI displays a code editor window titled "Log - (Untitled)" containing the following SAS code:

```
10 PROC SGplot DATA = Olympic1500;  
11 LOESS X = Year Y = Men / NOMARKERS CLM NOLEGCLM;  
12 REG X = Year Y = Men;  
13 LABEL Men = 'Time in Seconds';  
14 TITLE 'Olympic Times for Men's 1500 Meter Run';  
15 RUN;
```

Below the code, the log window shows the execution results:

```
NOTE: Writing HTML Body file: sashtml.htm  
NOTE: PROCEDURE SGplot used (Total process time):  
      real time           3.34 seconds  
      cpu time            0.67 seconds  
NOTE: There were 28 observations read from the data set WORK.OLYMPIC1500.
```

The bottom of the window shows the command window with the same code and the status bar indicating "Autosave complete" and "C:\Users\Susan Slaughter Ln 2, Col 18".

SAS

The SPSS GUI displays a data editor window with the following data:

position	agerang	yrsscale	hourwage	
1	1	1	2	13.74
2	0	1	2	16.44
3	0	1	3	21.39
4	1	1	1	11.38
5	0	1	3	21.56
6	0	1	1	18.12
7	1	1	3	13.14
8	0	1	1	24.73
9	0	1	2	15.70
10	1	1	1	18.94
11	0	1	1	25.45
12	0	1	1	19.71
13	1	1	2	21.14
14	0	1	2	20.53
15	0	1	2	20.83
16	1	1	2	16.81
17	0	1	2	17.59
18	0	1	3	18.73
19	1	1	2	14.77
20	0	1	3	19.36
21	0	1	2	17.03
22	1	1	3	.
23	0	1	3	20.67
24	0	1	2	19.41
25	1	1	3	20.22
26	0	1	1	20.23
27	0	1	3	16.48
28	1	1	3	12.27
29	0	1	3	23.51
30	0	1	2	17.67
31	1	1	2	11.20
32	0	1	1	20.44

The right side of the SPSS GUI shows the "Univariate Analysis of Variance" results for the dependent variable "Hourly Salary":

Source	Type III Sum of Squares	df	Mean Square	F	Sig.
Corrected Model	5701.240 <sup>a</sup>	11	518.295	36.709	.000
Intercept	758118.337	1	758118.337	53695.262	.000
position	2051.609	1	2051.609	145.309	.000
yrsscale	2872.222	5	574.444	40.686	.000
position * yrsscale	118.065	5	23.613	1.672	.138
Error	40930.707	2899	14.119		
Total	1212879.42	2911			
Corrected Total	46631.948	2910			

The bottom right of the SPSS GUI shows a "Profile Plots" window titled "Estimated Marginal Means of Hourly Salary". The plot shows the estimated marginal means for different years of experience (5 or less, 6-10, 11-15, 16-20, 21-35, 36 or more) for two nurse types: Hospital (blue bars) and Office (red bars). The observed grand mean is shown as a horizontal line. Error bars represent 95% confidence intervals.

SPSS

# Differences between Matlab and R

## 1. Open source:

- R is an open source while Matlab is not open source

## 2. Speed:

- R is slower than Matlab

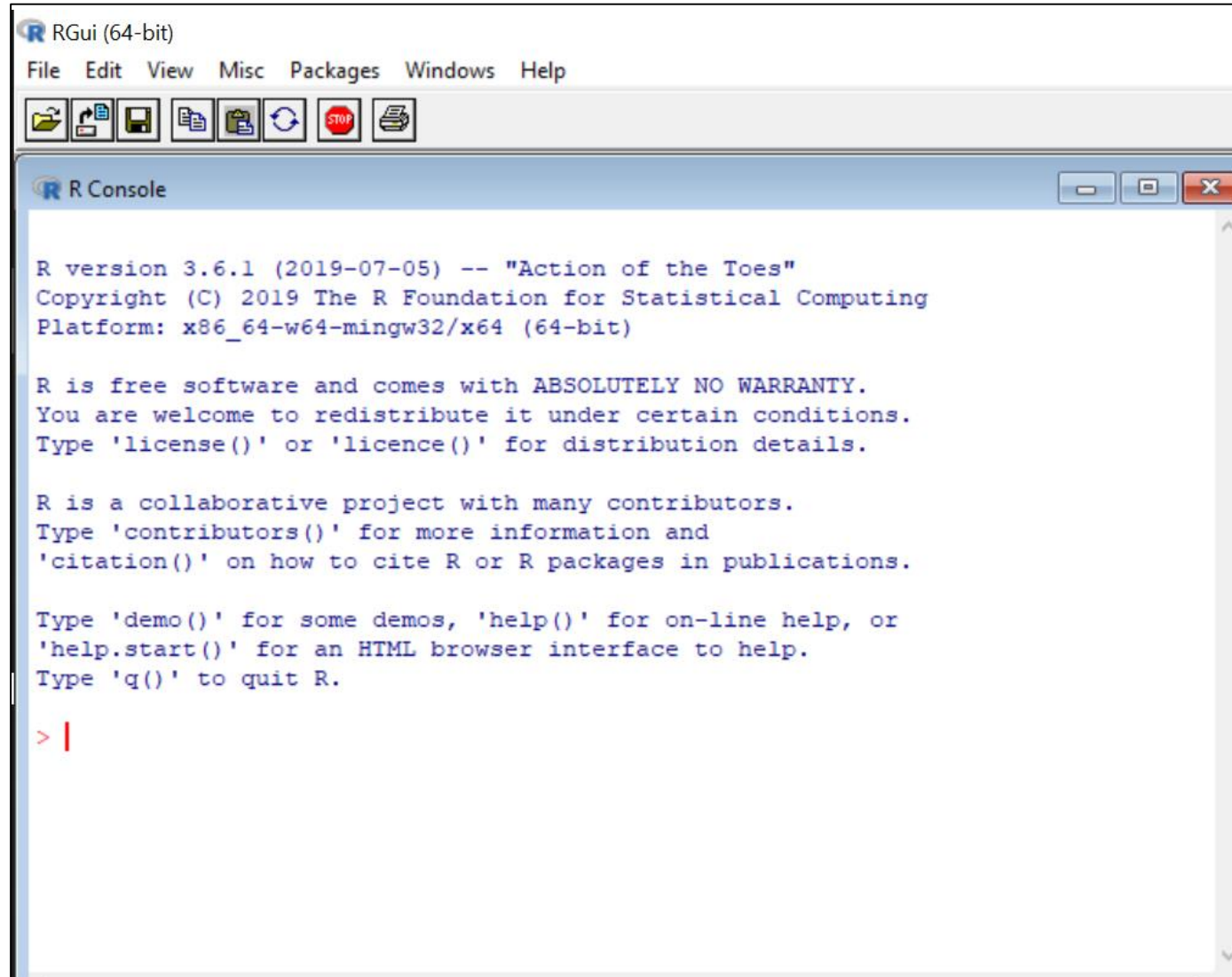
## 3. Functionality:

- R is mainly used for statistical analysis.
- Matlab is used for performing various engineering applications like image processing.

## 4. Ease of use :

- R follows programming language syntax.
- Matlab is easy to program as multiple toolbox are available

# R GUI



**Less fancy and no frills, but free !**

# Why to learn R ?

- **Since it is free and open-source.**
- **R consist of wide selection of additional libraries.**
- **Main library repositories CRAN and Bioconductor.**
- **A massive set of packages for statistical modelling, machine learning, visualization, and importing and manipulating data.**
- **Powerful tools for communicating your results.**
- **Deep-seated language support for data analysis. This includes features like missing values, data frames, and vectorization.**



# Why to learn R ?

- **Cutting edge tools.** Researchers in statistics and machine learning will often publish an R package to accompany their articles. This means immediate access to the very latest statistical techniques and implementations.
- **RMarkdown** makes it easy to turn your results into HTML files, PDFs, Word documents, PowerPoint presentations, dashboards and more.
- **Shiny** allows you to make beautiful interactive apps without any knowledge of HTML or javascript.
- **RStudio**, the IDE, provides an integrated development environment, tailored to the needs of data science, interactive data analysis, and statistical programming.
- **R** can connect easily to high-performance programming languages like C, Fortran, and C++.

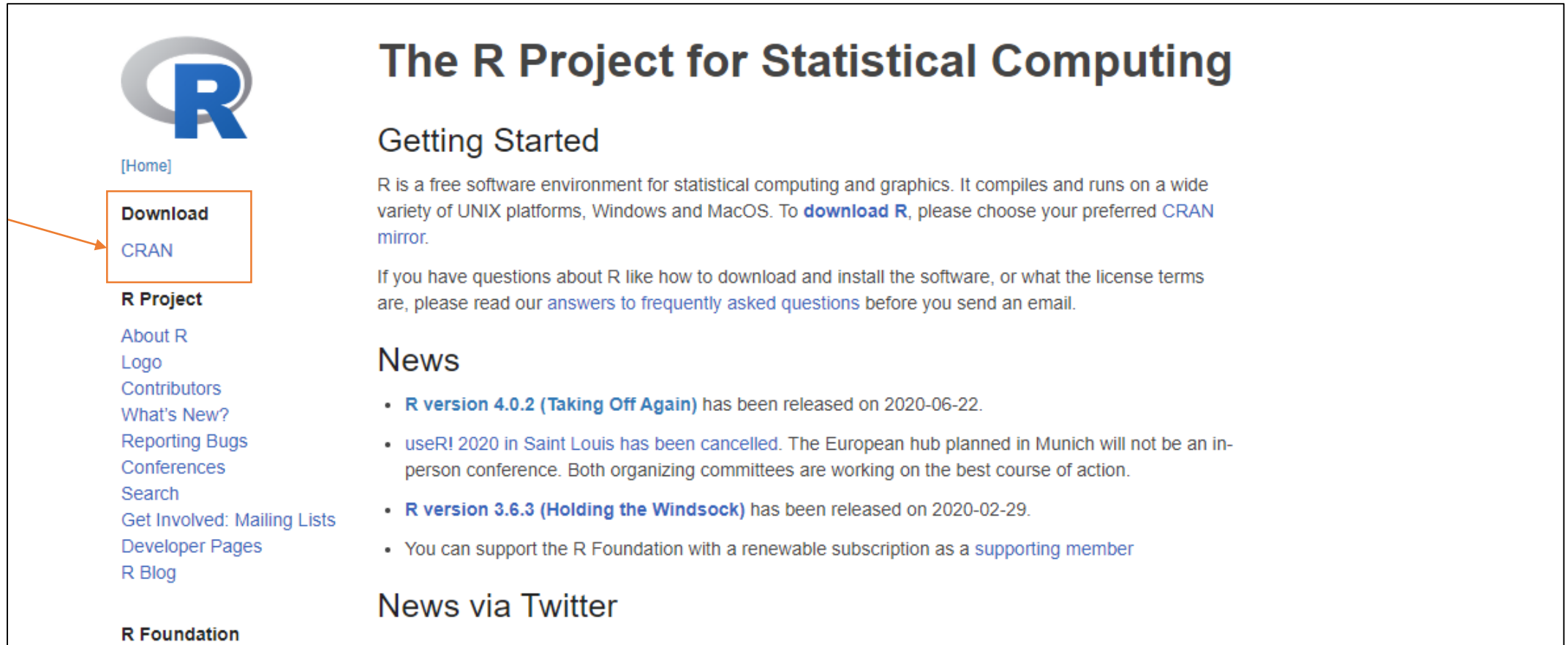
# Why to learn R ?

- **R's metaprogramming capabilities allow you to write magically succinct and concise functions.**
- **Excellent environment to handle ggplot2, dplyr, data.table, and more.**
- **Compared to other programming languages, the R community is more focused on results than processes.**

# Install R

<https://www.r-project.org/>

and do the following (assuming you work on a windows computer)



The screenshot shows the R Project website. On the left is a navigation menu with the R logo at the top. The 'Download CRAN' link is highlighted with an orange box and an arrow pointing to it. The main content area features the title 'The R Project for Statistical Computing', a 'Getting Started' section with introductory text, a 'News' section with a list of recent releases, and a 'News via Twitter' section. The footer includes the 'R Foundation' logo.

**The R Project for Statistical Computing**

Getting Started

R is a free software environment for statistical computing and graphics. It compiles and runs on a wide variety of UNIX platforms, Windows and MacOS. To **download R**, please choose your preferred [CRAN mirror](#).

If you have questions about R like how to download and install the software, or what the license terms are, please read our [answers to frequently asked questions](#) before you send an email.

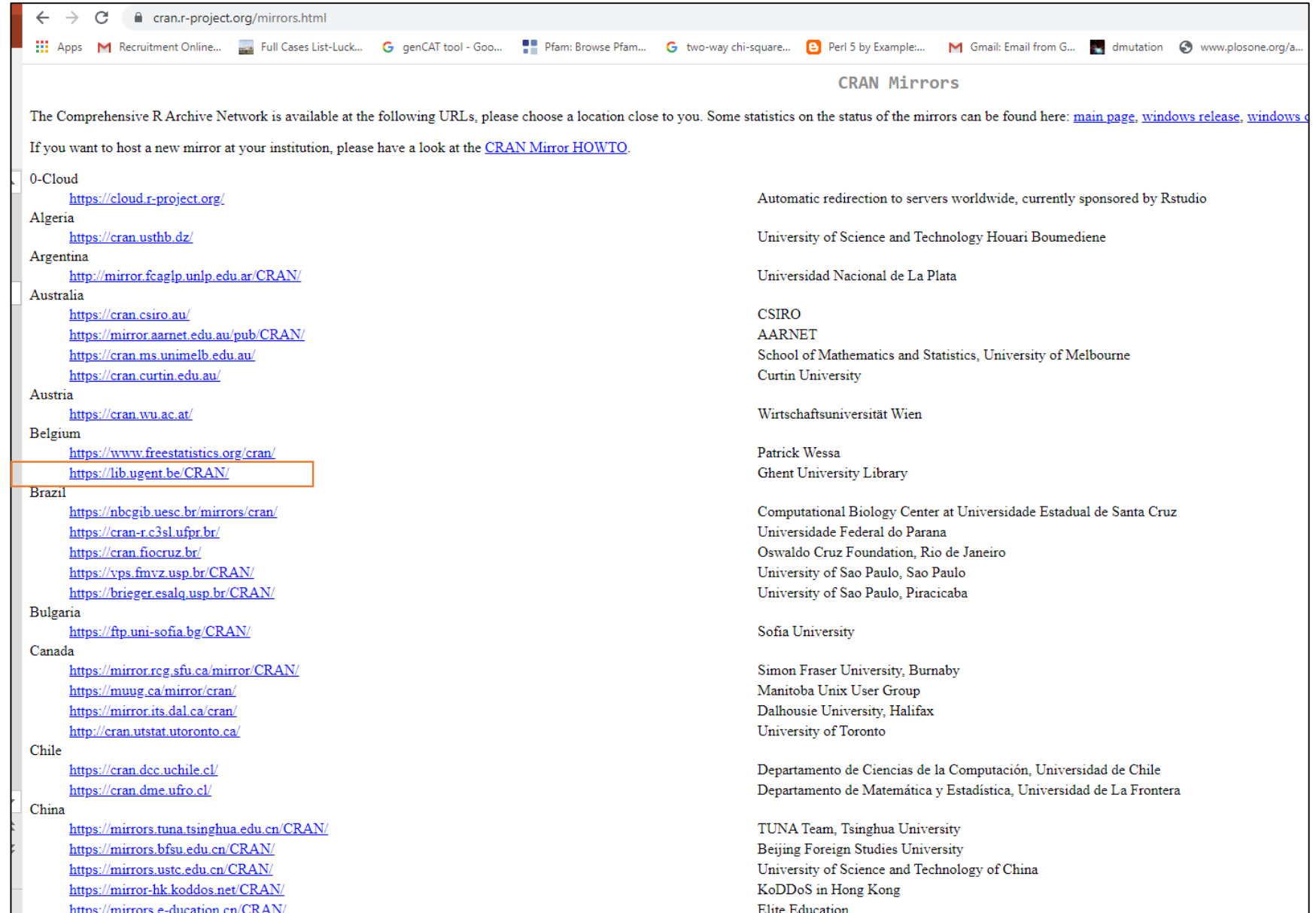
News

- **R version 4.0.2 (Taking Off Again)** has been released on 2020-06-22.
- [useR! 2020 in Saint Louis has been cancelled](#). The European hub planned in Munich will not be an in-person conference. Both organizing committees are working on the best course of action.
- **R version 3.6.3 (Holding the Windsock)** has been released on 2020-02-29.
- You can support the R Foundation with a renewable subscription as a [supporting member](#)

News via Twitter

R Foundation

# Choose download site



The screenshot shows the CRAN Mirrors page with the following content:

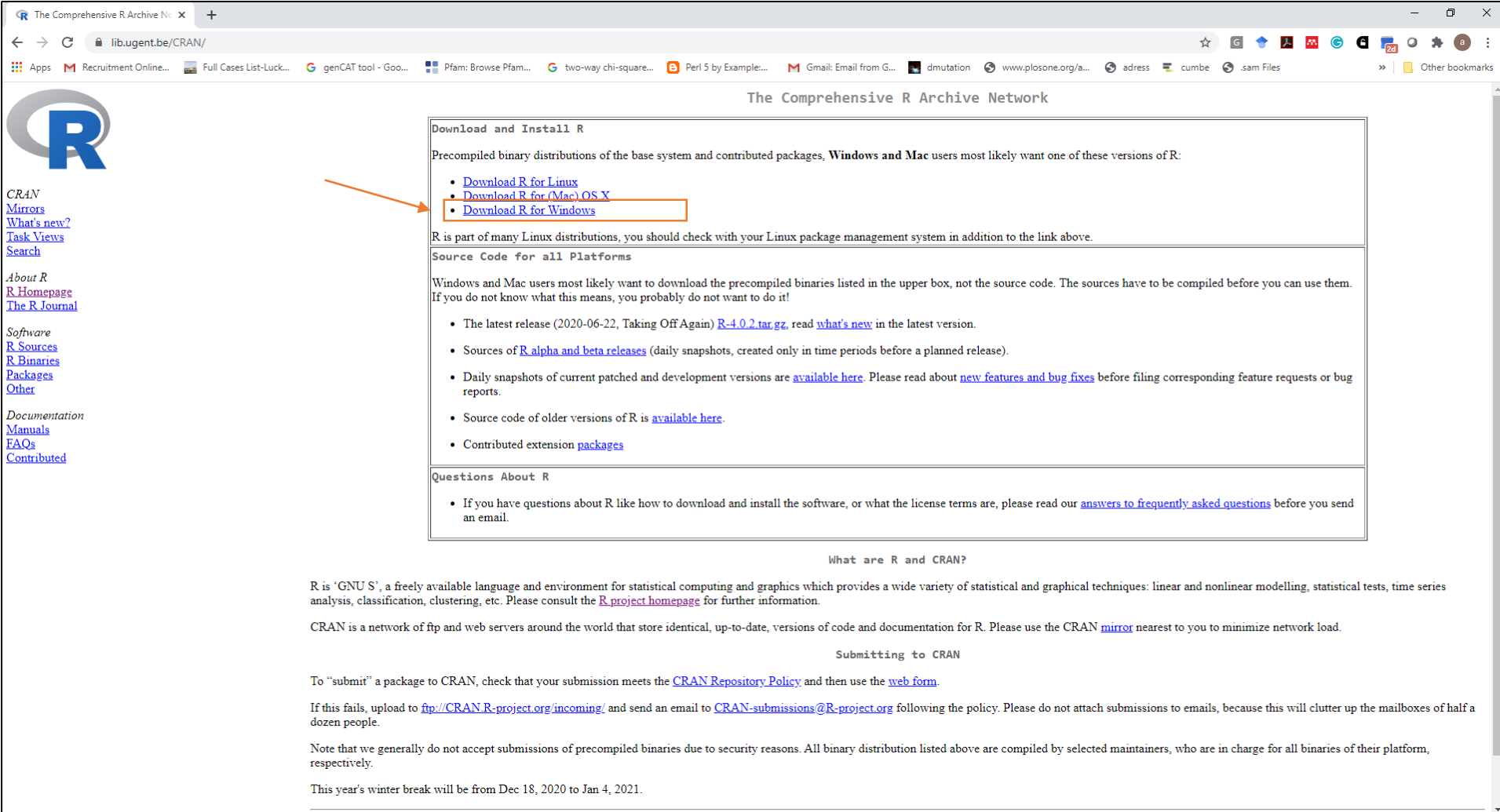
CRAN Mirrors

The Comprehensive R Archive Network is available at the following URLs, please choose a location close to you. Some statistics on the status of the mirrors can be found here: [main page](#), [windows release](#), [windows c](#)

If you want to host a new mirror at your institution, please have a look at the [CRAN Mirror HOWTO](#).

0-Cloud	<a href="https://cloud.r-project.org/">https://cloud.r-project.org/</a>	Automatic redirection to servers worldwide, currently sponsored by Rstudio
Algeria	<a href="https://cran.usthb.dz/">https://cran.usthb.dz/</a>	University of Science and Technology Houari Boumediene
Argentina	<a href="http://mirror.fcaglp.unlp.edu.ar/CRAN/">http://mirror.fcaglp.unlp.edu.ar/CRAN/</a>	Universidad Nacional de La Plata
Australia	<a href="https://cran.csiro.au/">https://cran.csiro.au/</a> <a href="https://mirror.aarnet.edu.au/pub/CRAN/">https://mirror.aarnet.edu.au/pub/CRAN/</a> <a href="https://cran.ms.unimelb.edu.au/">https://cran.ms.unimelb.edu.au/</a> <a href="https://cran.curtin.edu.au/">https://cran.curtin.edu.au/</a>	CSIRO AARNET School of Mathematics and Statistics, University of Melbourne Curtin University
Austria	<a href="https://cran.wu.ac.at/">https://cran.wu.ac.at/</a>	Wirtschaftsuniversität Wien
Belgium	<a href="https://www.freeststatistics.org/cran/">https://www.freeststatistics.org/cran/</a> <a href="https://lib.ugent.be/CRAN/">https://lib.ugent.be/CRAN/</a>	Patrick Wessa Ghent University Library
Brazil	<a href="https://mbcgib.uesc.br/mirrors/cran/">https://mbcgib.uesc.br/mirrors/cran/</a> <a href="https://cran-r.c3sl.ufpr.br/">https://cran-r.c3sl.ufpr.br/</a> <a href="https://cran.fiocruz.br/">https://cran.fiocruz.br/</a> <a href="https://vps.fmvz.usp.br/CRAN/">https://vps.fmvz.usp.br/CRAN/</a> <a href="https://brieger.esalq.usp.br/CRAN/">https://brieger.esalq.usp.br/CRAN/</a>	Computational Biology Center at Universidade Estadual de Santa Cruz Universidade Federal do Parana Oswaldo Cruz Foundation, Rio de Janeiro University of Sao Paulo, Sao Paulo University of Sao Paulo, Piracicaba
Bulgaria	<a href="https://ftp.uni-sofia.bg/CRAN/">https://ftp.uni-sofia.bg/CRAN/</a>	Sofia University
Canada	<a href="https://mirror.rcg.sfu.ca/mirror/CRAN/">https://mirror.rcg.sfu.ca/mirror/CRAN/</a> <a href="https://muug.ca/mirror/cran/">https://muug.ca/mirror/cran/</a> <a href="https://mirror.its.dal.ca/cran/">https://mirror.its.dal.ca/cran/</a> <a href="http://cran.utstat.utoronto.ca/">http://cran.utstat.utoronto.ca/</a>	Simon Fraser University, Burnaby Manitoba Unix User Group Dalhousie University, Halifax University of Toronto
Chile	<a href="https://cran.dcc.uchile.cl/">https://cran.dcc.uchile.cl/</a> <a href="https://cran.dme.ufro.cl/">https://cran.dme.ufro.cl/</a>	Departamento de Ciencias de la Computación, Universidad de Chile Departamento de Matemática y Estadística, Universidad de La Frontera
China	<a href="https://mirrors.tuna.tsinghua.edu.cn/CRAN/">https://mirrors.tuna.tsinghua.edu.cn/CRAN/</a> <a href="https://mirrors.bfsu.edu.cn/CRAN/">https://mirrors.bfsu.edu.cn/CRAN/</a> <a href="https://mirrors.ustc.edu.cn/CRAN/">https://mirrors.ustc.edu.cn/CRAN/</a> <a href="https://mirror-hk.koddos.net/CRAN/">https://mirror-hk.koddos.net/CRAN/</a> <a href="https://mirrors.e-ducation.cn/CRAN/">https://mirrors.e-ducation.cn/CRAN/</a>	TUNA Team, Tsinghua University Beijing Foreign Studies University University of Science and Technology of China KoDDoS in Hong Kong Elite Education

# Choose Windows as target operation system



The screenshot shows the CRAN website with the following content:

## The Comprehensive R Archive Network

### Download and Install R

Precompiled binary distributions of the base system and contributed packages, **Windows and Mac** users most likely want one of these versions of R:

- [Download R for Linux](#)
- [Download R for \(Mac\) OS X](#)
- [Download R for Windows](#)

R is part of many Linux distributions, you should check with your Linux package management system in addition to the link above.

### Source Code for all Platforms

Windows and Mac users most likely want to download the precompiled binaries listed in the upper box, not the source code. The sources have to be compiled before you can use them. If you do not know what this means, you probably do not want to do it!

- The latest release (2020-06-22, Taking Off Again) [R-4.0.2.tar.gz](#), read [what's new](#) in the latest version.
- Sources of [R alpha and beta releases](#) (daily snapshots, created only in time periods before a planned release).
- Daily snapshots of current patched and development versions are [available here](#). Please read about [new features and bug fixes](#) before filing corresponding feature requests or bug reports.
- Source code of older versions of R is [available here](#).
- Contributed extension [packages](#)

### Questions About R

- If you have questions about R like how to download and install the software, or what the license terms are, please read our [answers to frequently asked questions](#) before you send an email.

### What are R and CRAN?

R is "GNU S", a freely available language and environment for statistical computing and graphics which provides a wide variety of statistical and graphical techniques: linear and nonlinear modelling, statistical tests, time series analysis, classification, clustering, etc. Please consult the [R project homepage](#) for further information.

CRAN is a network of ftp and web servers around the world that store identical, up-to-date, versions of code and documentation for R. Please use the CRAN [mirror](#) nearest to you to minimize network load.

### Submitting to CRAN

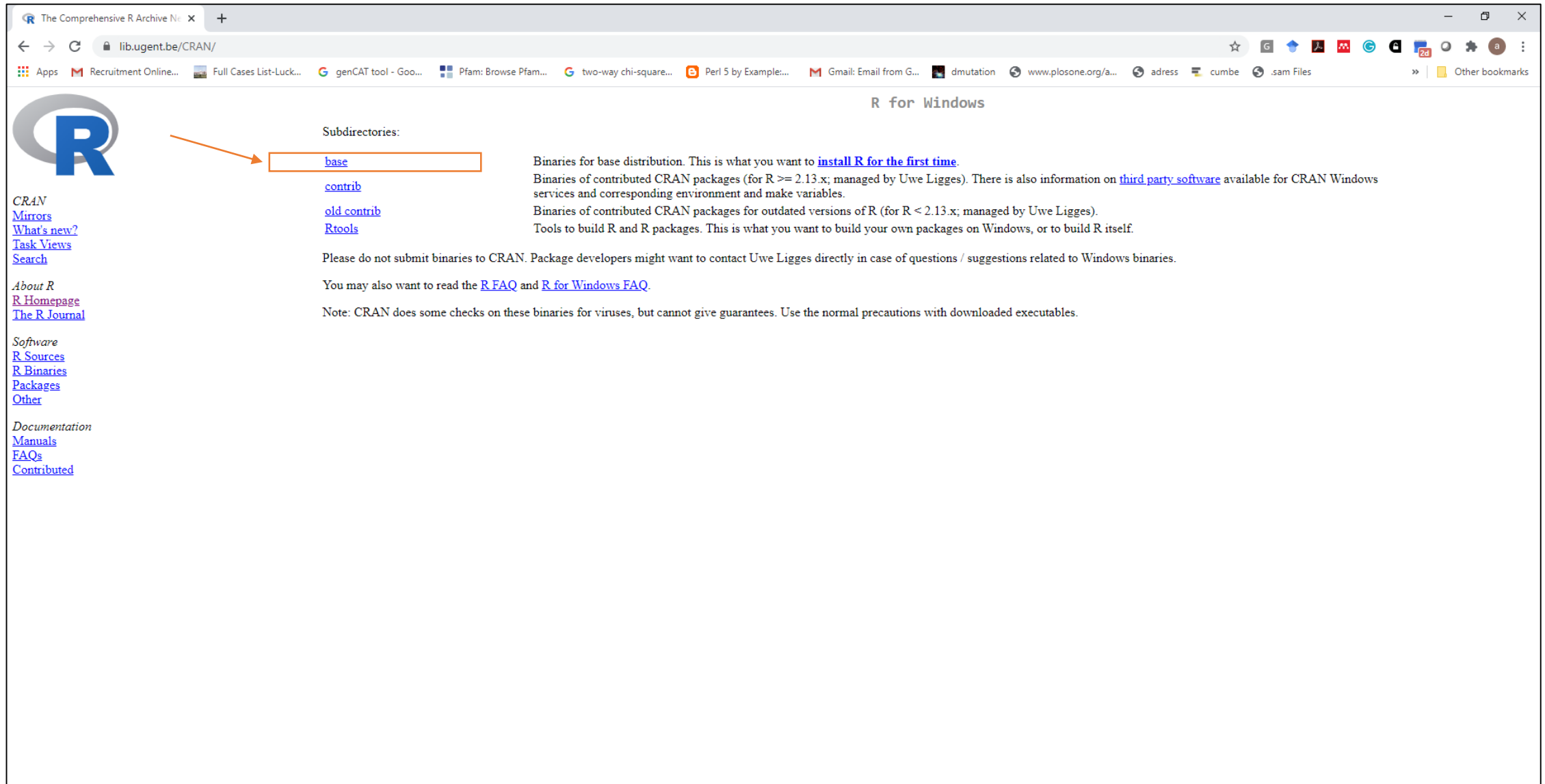
To "submit" a package to CRAN, check that your submission meets the [CRAN Repository Policy](#) and then use the [web form](#).

If this fails, upload to <ftp://CRAN.R-project.org/incoming/> and send an email to [CRAN-submissions@R-project.org](mailto:CRAN-submissions@R-project.org) following the policy. Please do not attach submissions to emails, because this will clutter up the mailboxes of half a dozen people.

Note that we generally do not accept submissions of precompiled binaries due to security reasons. All binary distribution listed above are compiled by selected maintainers, who are in charge for all binaries of their platform, respectively.

This year's winter break will be from Dec 18, 2020 to Jan 4, 2021.

# Click base



The screenshot shows a web browser window with the address bar at `lib.ugent.be/CRAN/`. The page title is "R for Windows". On the left side, there is a navigation menu with links for "CRAN", "Mirrors", "What's new?", "Task Views", "Search", "About R", "R Homepage", "The R Journal", "Software", "R Sources", "R Binaries", "Packages", "Other", "Documentation", "Manuals", "FAQs", and "Contributed". The main content area is titled "Subdirectories:" and lists four links: "base", "contrib", "old\_contrib", and "Rtools". The "base" link is highlighted with an orange rectangular box, and an orange arrow points from the CRAN logo to this box. To the right of the "base" link, there is a paragraph of text: "Binaries for base distribution. This is what you want to [install R for the first time](#)." Below this, there are two more paragraphs: "Binaries of contributed CRAN packages (for R >= 2.13.x; managed by Uwe Ligges). There is also information on [third party software](#) available for CRAN Windows services and corresponding environment and make variables." and "Binaries of contributed CRAN packages for outdated versions of R (for R < 2.13.x; managed by Uwe Ligges).". Below these, there is a paragraph: "Tools to build R and R packages. This is what you want to build your own packages on Windows, or to build R itself." At the bottom of the subdirectory list, there are two more paragraphs: "Please do not submit binaries to CRAN. Package developers might want to contact Uwe Ligges directly in case of questions / suggestions related to Windows binaries." and "You may also want to read the [R FAQ](#) and [R for Windows FAQ](#)." At the very bottom of the subdirectory list, there is a note: "Note: CRAN does some checks on these binaries for viruses, but cannot give guarantees. Use the normal precautions with downloaded executables."

# Click Download R 4.0.2

The screenshot shows a web browser window with the address bar displaying `lib.ugent.be/CRAN/`. The page title is "R-4.0.2 for Windows (32/64 bit)". The R logo is on the left. A navigation menu includes links for "CRAN", "Mirrors", "What's new?", "Task Views", "Search", "About R", "R Homepage", "The R Journal", "Software", "R Sources", "R Binaries", "Packages", "Other", "Documentation", "Manuals", "FAQs", and "Contributed". The main content area features a large button labeled "Download R 4.0.2 for Windows (84 megabytes, 32/64 bit)", which is highlighted with an orange box and an orange arrow. Below this button are links for "Installation and other instructions" and "New features in this version". A paragraph of text explains how to verify the downloaded package using `md5sum` and provides links for "graphical" and "command line versions". A "Frequently asked questions" section lists three questions: "Does R run under my version of Windows?", "How do I update packages in my previous version of R?", and "Should I run 32-bit or 64-bit R?". Below this is a note to refer to the "R FAQ" and "R Windows FAQ". An "Other builds" section lists three items: "Patches to this release are incorporated in the r-patched snapshot build.", "A build of the development version (which will eventually become the next major release of R) is available in the r-devel snapshot build.", and "Previous releases". A note to webmasters provides a stable link: `<CRAN MIRROR>/bin/windows/base/release.html`. The footer indicates "Last change: 2020-06-22".

Download the .exe file and run it (choose default answers for all questions) 15

# Install Rstudio


<https://rstudio.com/products/rstudio/download/>

## Download RStudio

### Choose Your Version

RStudio is a set of integrated tools designed to help you be more productive with R. It includes a console, syntax-highlighting editor that supports direct code execution, and a variety of robust tools for plotting, viewing history, debugging and managing your workspace.

[LEARN MORE ABOUT RSTUDIO FEATURES](#)



RStudio's new solution for every professional data science team. RStudio Team includes RStudio Server Pro, RStudio Connect and RStudio Package Manager.

[LEARN MORE](#)

RStudio Desktop Open Source License <b>Free</b>	RStudio Desktop Commercial License <b>\$995</b> /year	RStudio Server Open Source License <b>Free</b>	RStudio Server Pro Commercial License <b>\$4,975</b> /year (5 Named Users)
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RStudio Desktop

Open Source License

**Free**

RStudio Desktop

Commercial License

**\$995**

/year

RStudio Server

Open Source License

**Free**

RStudio Server Pro

Commercial License

**\$4,975**

/year

(5 Named Users)

**DOWNLOAD**

**BUY**

**DOWNLOAD**

**BUY**

RStudio Desktop 1.3.1073 - [Release Notes](#)

1. Install R. RStudio requires R 3.0.1+.
2. Download RStudio Desktop. Recommended for your system:



**DOWNLOAD RSTUDIO FOR WINDOWS**  
1.3.1073 | 171.62MB

Requires Windows 10/8/7 (64-bit)



# Rstudio layout

The screenshot displays the RStudio environment with the following components:

- Source Editor:** Contains a script named 'hello.R' with the following code:

```
1 # Hello, world!  
2 #  
3 # This is an example function named 'hello'  
4 # which prints 'Hello, world!'.  
5 #  
6 # You can learn more about package authoring with RStudio at:  
7 #  
8 # http://r-pkgs.had.co.nz/  
9 #  
10 # Some useful keyboard shortcuts for package authoring:  
11 #  
12 # Install Package:      'Ctrl + Shift + B'  
13 # Check Package:       'Ctrl + Shift + E'  
14 # Test Package:        'Ctrl + Shift + T'  
15 #  
16 hello <- function() {  
17   print("Hello, world!")  
18 }
```
- Environment Panel:** Shows 'Global Environment' with the message 'Environment is empty'.
- Files Panel:** Displays the file explorer for the path 'C:\RESEARCH\OV\DAI'. The file list is as follows:

Name	Size	Type
..		Folder
.gitignore	44 B	File
.Rbuildignore	30 B	File
.Rhistory	111 B	File
DAI.Rproj	376 B	File
DESCRIPTION	376 B	File
man		Folder
NAMESPACE	32 B	File
R		Folder
- Console:** Shows the R startup message:

```
You are welcome to redistribute it under certain conditions.  
Type 'license()' or 'licence()' for distribution details.  
  
R is a collaborative project with many contributors.  
Type 'contributors()' for more information and  
'citation()' on how to cite R or R packages in publications.  
  
Type 'demo()' for some demos, 'help()' for on-line help, or  
'help.start()' for an HTML browser interface to help.  
Type 'q()' to quit R.  
  
> |
```

# RStudio

- **Bottom left : console window . Here you can type simple commands after the > prompt and R will execute your command. This is the most important window, because this is where R actually does stuff.**
- **Top left : editor window (also called script window). Collections of commands (scripts) can be edited and saved.**
- **Top right : workspace/history window. In the workspace window, you can see which data and values R has in its memory. You can view and edit the values by clicking on them. This history window shows what has been typed before.**
- **Bottom right : files/plots/packages/help window. Here you can open files, view plots (also previous plots), install and load packages and use the help function.**

**Use R or RStudio**

**Lets use it!**

# Operators and DATA TYPES

# Variables/Operators

- Variables store one element

```
x <- 25
```

Here x variable is assigned value 25

- Check value assigned to the variable x

```
> x  
[1] 25
```

- Basic mathematical operators that could be applied to variables : (+), (-), (-), (\*)
- Use parenthesis to obtain desired sequence of mathematical operations

## Calculator

R can be used as a calculator. You can just type your equation in the command windows after the > :

```
> 10 + 20
```

## Workspace

You can also give numbers a name. By doing so, they become so-called variables which can be used later. For example, you can type in the command window:

```
> a = 4
```

- You can also ask R what a is (just type a ENTER in the command window):

```
> a
```

- Or do calculations with a :

```
> a * 5
```

- To remove all variables from R's memory, type

```
> rm(list=ls())
```



# Concatenation function

**c()** ,

```
>x <- c(1,2,3,4,5)
>x
>y <- c("a", "b", "c", "d")
>y
```

# Vectors

- **Vectors have only 1 dimension and represent enumerated sequence of data. They can also store variables**

```
> v1 <- c(1,2,3,4)
> mean (v1)
```

- **The elements of a vector are specified/modified with braces (e.g. [number])**

```
> v1[1] <- 48
v1
[1] 48 2 3 4
```

# Logical operators

- These operators mostly work on vectors, matrices and other data types.
- Type of data is not important, the same operators are used for numeric and character data type

Operator	Description
<	less than
<=	less than or equal to
>	greater than
>=	greater than or equal to
==	exactly equal to
!=	not equal to

# R workspace

- Display all workplace objects (variables, vectors etc.) via ls():

```
> a= 10  
> b =20  
> ls()  
[1] "a" "b"
```

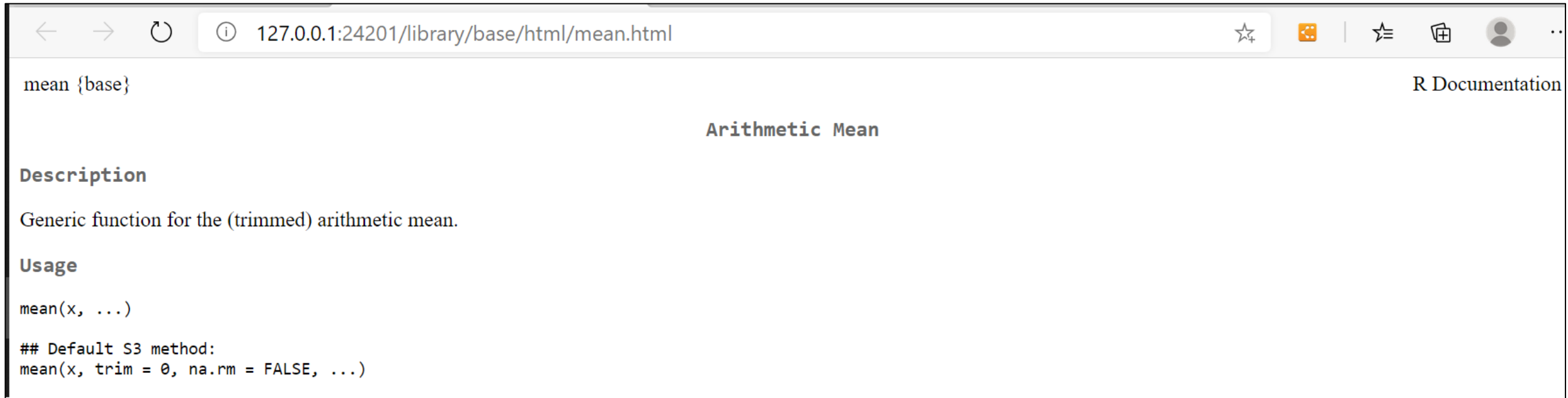
- Useful tip : to save workplace and restore from a file use:

```
>save.image(file="workspace.rda")  
>load(file="workspace.rda")
```

# Information of function

- Any function in R has help information
- To invoke help use `? Sign` and `help()`:

```
> ? mean  
> help(mean)
```



```
mean {base} R Documentation  
  
Arithmetic Mean  
  
Description  
Generic function for the (trimmed) arithmetic mean.  
  
Usage  
mean(x, ...)  
  
## Default S3 method:  
mean(x, trim = 0, na.rm = FALSE, ...)
```

# Data types

- Data could be of 3 basic data types:
- numeric
- Character
- logical

**Numeric variable type :**

```
> x <- 1  
> mode(x)  
[1] "numeric"
```

## logical variable type :

```
> y <- 3 < 4
> y
[1] TRUE
> mode(y)
[1] "logical"
```

## Character variable type :

```
> a = "bioinfo"
> mode(a)
[1] "character"
```

# Data objects

- The main data objects in R :
  - Matrices (single data type)
  - Data frames (supports various data types)
  - Lists (contains set of vectors)
- Matrices are 2D objects (rows/columns) :

```
> m <- matrix(0,2,3)
```

```
> m
```

```
      [,1] [,2] [,3]  
[1,]  0   0   0  
[2,]  0   0   0
```



# List

- List contain various vectors. Each vectors in the list can be accessed by double braces `[[number]]`

```
> x <- c(1,2,3,4)
> y <- c(2,3,4)
> L1 <- list(x,y)
> L1
[[1]]
[1] 1 2 3 4
[[2]]
[1] 2 3 4
```

# Data frames

- Data frames are similar to matrices but can contain various data types

```
> x <- c(1,5,10)
> y <- c("A","B","C")
> z <- data.frame(x,y)
> z
  x y
1 1 A
2 5 B
3 10 C
```

# **RStudio : Advance Feature**

# R markdown

- Markdown is a simple formatting language designed to make authoring content easy for everyone.
- Rather than write in complex markup code (e.g. HTML or LaTeX), you write in plain text with formatting cues.

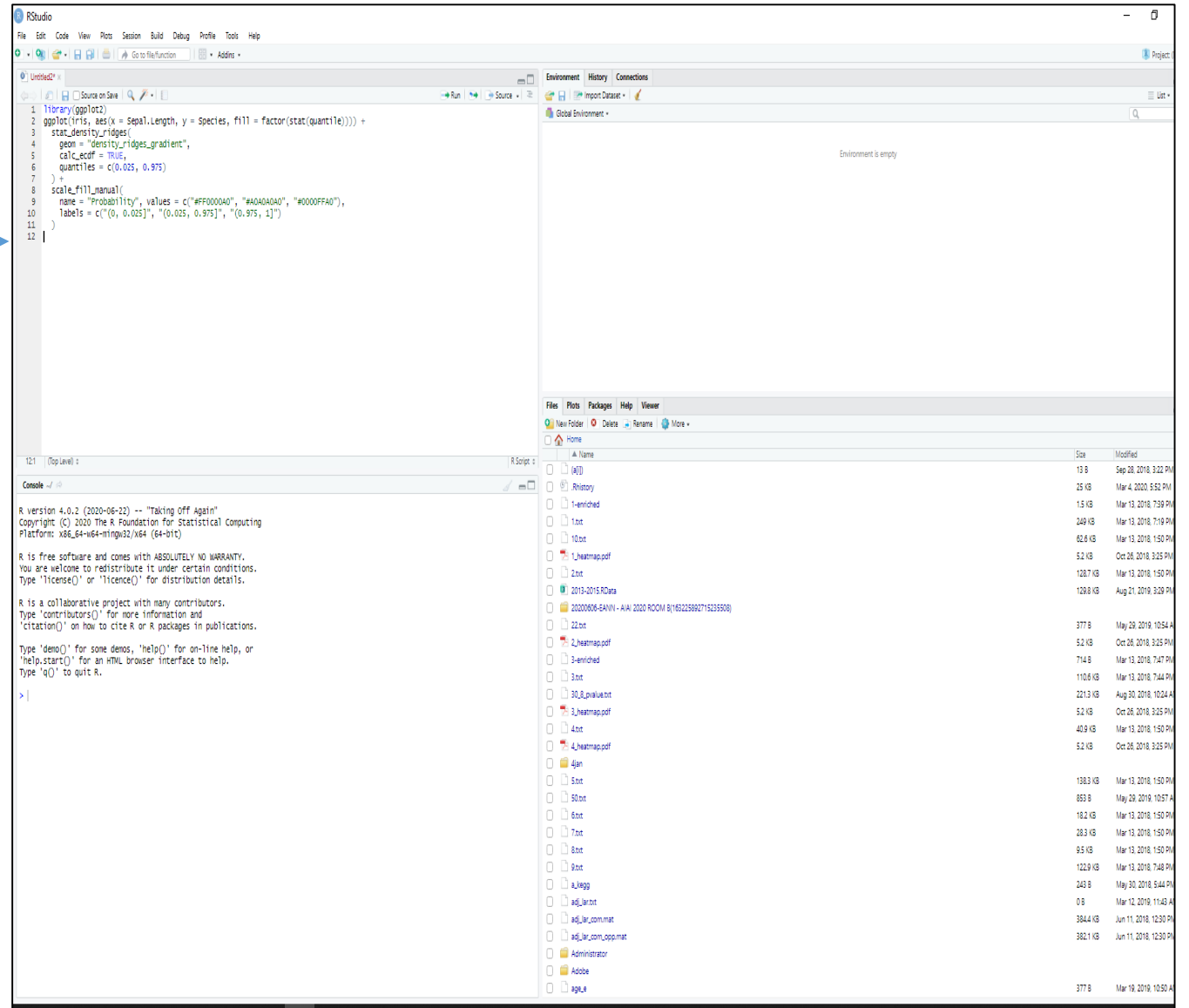
```
install.packages("rmarkdown")
```

```
install.packages("ggplot2")
```

```
install.packages("ggridges")
```

```
library(ggplot2)
library(ggribbles)
```

```
ggplot(iris, aes(x = Sepal.Length, y =
Species, fill = factor(stat(quantile)))) +
stat_density_ridges(
  geom = "density_ridges_gradient",
  calc_ecdf = TRUE,
  quantiles = c(0.025, 0.975)
) +
scale_fill_manual(
  name = "Probability", values =
c("#FF0000A0", "#A0A0A0A0",
"#0000FFA0"),
  labels = c("(0, 0.025]", "(0.025, 0.975]",
"(0.975, 1]")
)
```



```
RStudio
File Edit Code View Plots Session Build Debug Profile Tools Help
[Icons] Go to File/Function [Add]
1 library(ggplot2)
2 ggplot(iris, aes(x = Sepal.Length, y = Species, fill = factor(stat(quantile)))) +
3   stat_density_ridges(
4     geom = "density_ridges_gradient",
5     calc_ecdf = TRUE,
6     quantiles = c(0.025, 0.975)
7   ) +
8   scale_fill_manual(
9     name = "Probability", values = c("#FF0000A0", "#A0A0A0A0", "#0000FFA0"),
10    labels = c("(0, 0.025]", "(0.025, 0.975]", "(0.975, 1]")
11  )
12 |

Environment History Connections
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File Plots Packages Help Viewer
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Home
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[Icons] 377 B May 29 2019 10:54 A
[Icons] 5.2 KB Oct 26 2018 3:25 PM
[Icons] 714 B Mar 13 2018 7:47 PM
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[Icons] 221.3 KB Aug 30 2018 10:24 A
[Icons] 5.2 KB Oct 26 2018 3:25 PM
[Icons] 40.9 KB Mar 13 2018 1:50 PM
[Icons] 5.2 KB Oct 26 2018 3:25 PM
[Icons] 198.3 KB Mar 13 2018 1:50 PM
[Icons] 853 B May 29 2019 10:57 A
[Icons] 18.2 KB Mar 13 2018 1:50 PM
[Icons] 28.3 KB Mar 13 2018 1:50 PM
[Icons] 9.5 KB Mar 13 2018 1:50 PM
[Icons] 122.9 KB Mar 13 2018 7:42 PM
[Icons] 249 B May 30 2018 5:44 PM
[Icons] 0 B Mar 12 2019 11:43 A
[Icons] 384.4 KB Jun 11 2018 12:30 PM
[Icons] 382.1 KB Jun 11 2018 12:30 PM
[Icons] Administrator
[Icons] Adobe
[Icons] agluE 377 B Mar 16 2019 10:50 A

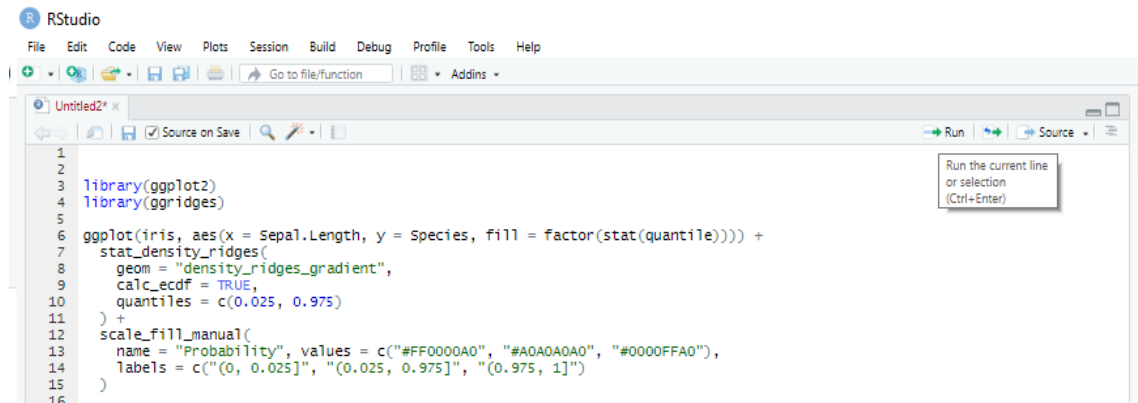
R version 4.0.2 (2020-06-22) -- "Taking Off Again"
Copyright (C) 2020 The R Foundation for Statistical Computing
Platform: x86_64-w64-mingw32/x64 (64-bit)

R is free software and comes with ABSOLUTELY NO WARRANTY.
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Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.

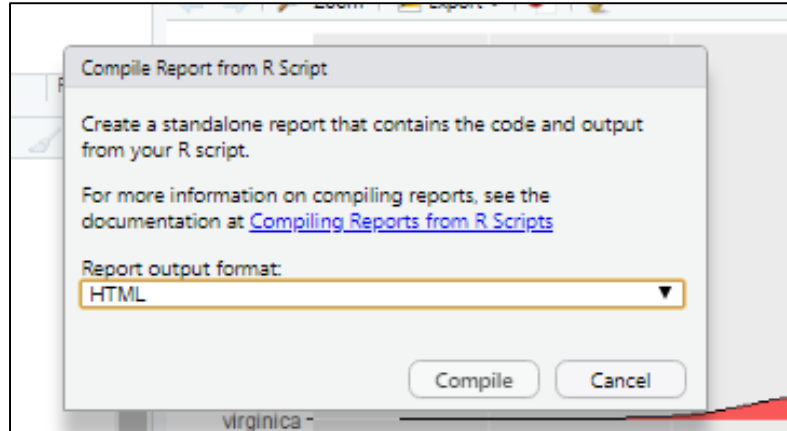
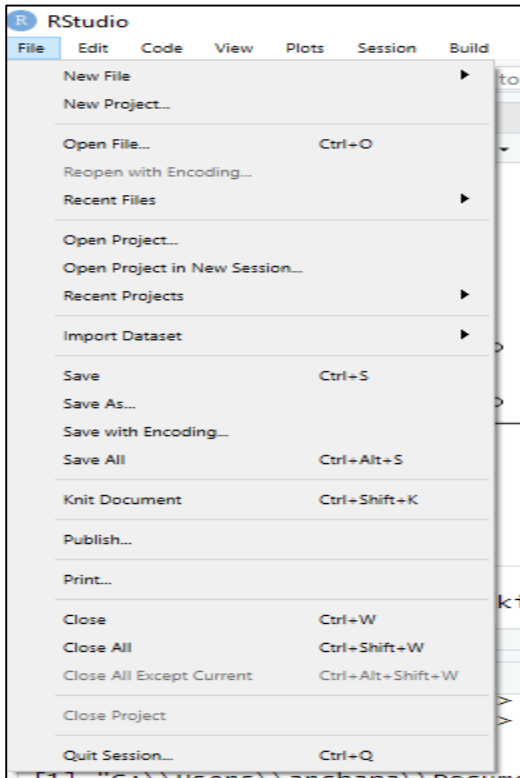
Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.
> |
```

- Run code

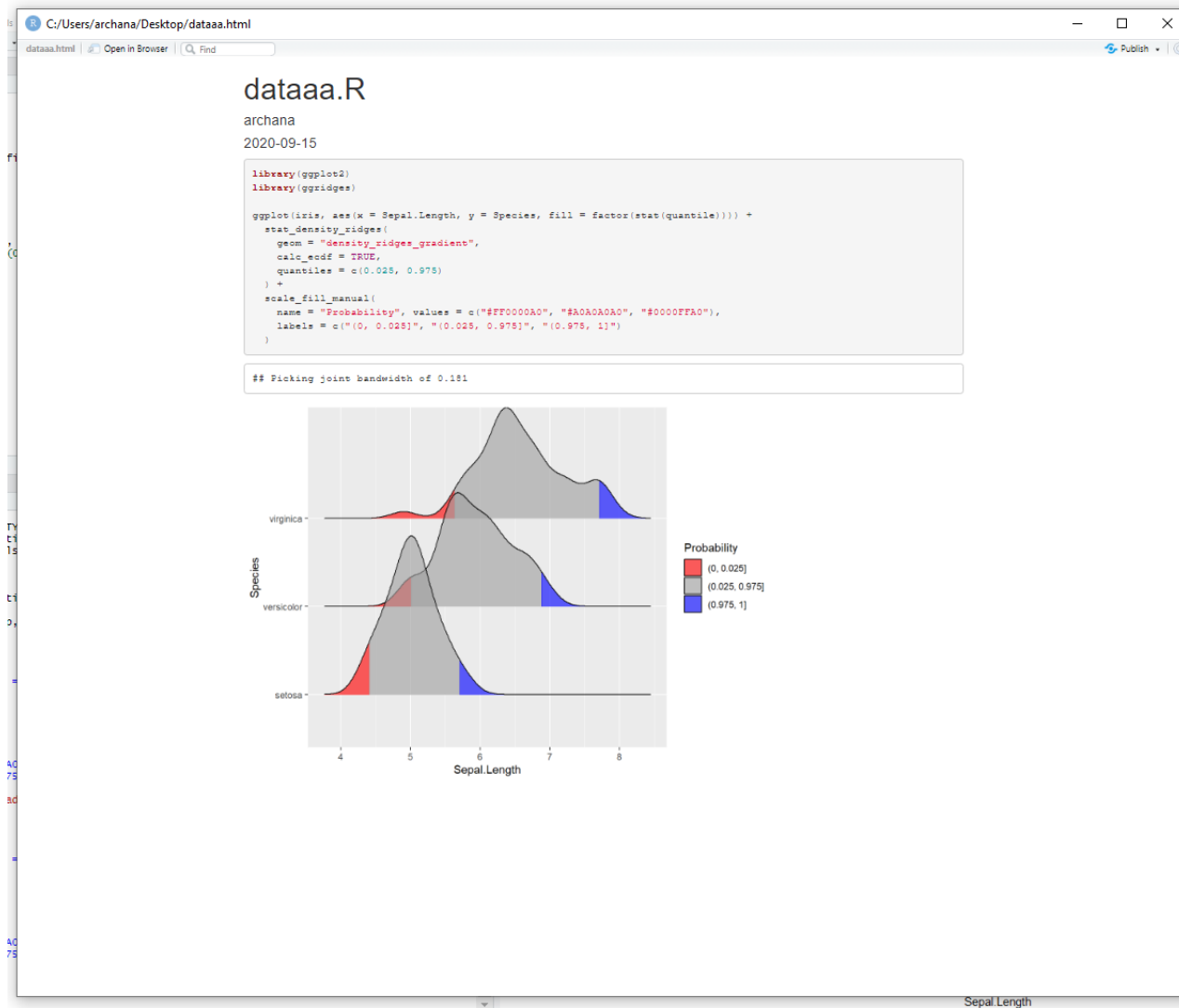


```
1  
2  
3 library(ggplot2)  
4 library(ggribes)  
5  
6 ggplot(iris, aes(x = Sepal.Length, y = Species, fill = factor(stat(quantile)))) +  
7   stat_density_ridges(  
8     geom = "density_ridges_gradient",  
9     calc_ecdf = TRUE,  
10    quantiles = c(0.025, 0.975)  
11  ) +  
12  scale_fill_manual(  
13    name = "Probability", values = c("#FF0000A0", "#A0A0A0A0", "#0000FFA0"),  
14    labels = c("(0, 0.025]", "(0.025, 0.975]", "(0.975, 1]")  
15  )  
16
```

- Click option File -> knit document



Report done



# Inbuilt functions in R

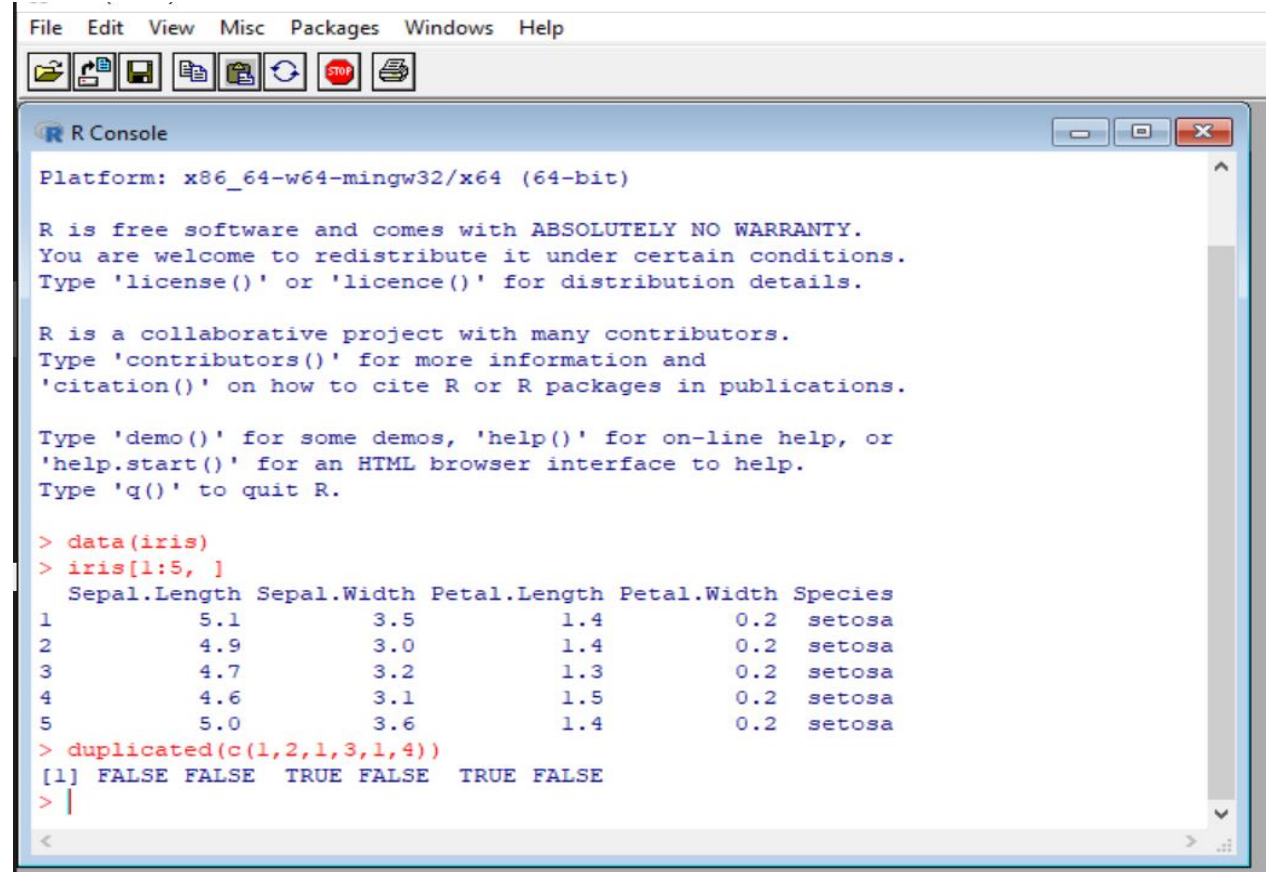


# deduplicated()function in R

- Duplicate data can be removed during analysis.
- It returns a logical vector that tells you whether the specified value is a duplicate of a previous value.

> duplicated(c(1,2,1,3,1,4))

**For all those values which are duplicate in the sample, true is returned.**



```
File Edit View Misc Packages Windows Help
R Console
Platform: x86_64-w64-mingw32/x64 (64-bit)

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Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.

Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.

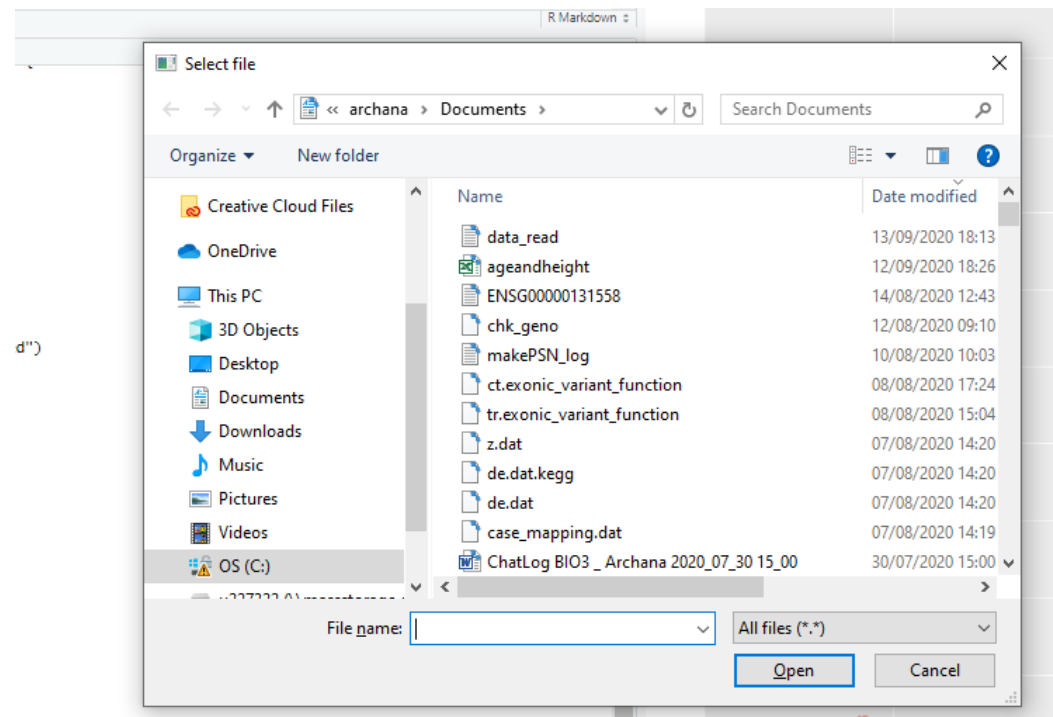
> data(iris)
> iris[1:5, ]
  Sepal.Length Sepal.Width Petal.Length Petal.Width Species
1           5.1           3.5           1.4           0.2  setosa
2           4.9           3.0           1.4           0.2  setosa
3           4.7           3.2           1.3           0.2  setosa
4           4.6           3.1           1.5           0.2  setosa
5           5.0           3.6           1.4           0.2  setosa

> duplicated(c(1,2,1,3,1,4))
[1] FALSE FALSE  TRUE FALSE  TRUE FALSE
> |
```

# Read data in R

- `file.choose()` to walk through your directory to select a file and load.
- In case you don't want header then set `header=FALSE`.

```
> data <- file.choose()
```



```
> data_R <- read.table(data)
```

```
> data_R
```

```
> data_R
```

```
      sample1 sample2 sample3 sample4 sample5 sample6  
gene1      3      1      3      1      3      1  
gene2      2      2      2      2      2      2  
gene3      1      7      1      7      1      7  
gene4      2      6      2      6      2      6  
gene5      3      3      3      3      3      3  
gene6      3      2      3      2      3      2  
gene7      2      2      2      2      2      2  
gene8      1      1      1      1      1      1
```

# Write data in R

- In R, we can write data frames easily to a file, using the `write.table()` command.
- The first argument refers to the data frame to be written to the output file, the second is the name of the output file.
- By default R will surround each entry in the output file by quotes, so we use `quote=F`

```
> write.table(x, file, sep = " ", row.names = TRUE, col.names = TRUE)
```

# na.omit() function in R

- Rows which have NA values can be removed using the na.omit() function as below:
- Download data from website

```
> read <- read.table(file="data_read2.txt")
> read
  sample1 sample2 sample3 sample4 sample5 sample6
gene1    3     1     3     1     3     1
gene2    2     2     2     2     2     2
gene3   NA     7     1     7     1     7
gene4    2     6     2     6     2     6
gene5    3     3     3     3     3     3
gene6   NA     2     3     2     3     2
gene7    2     2     2     2     2     2
gene8   NA     1     1     1     1     1
> na.omit(read)
  sample1 sample2 sample3 sample4 sample5 sample6
gene1    3     1     3     1     3     1
gene2    2     2     2     2     2     2
gene4    2     6     2     6     2     6
gene5    3     3     3     3     3     3
gene7    2     2     2     2     2     2
```

# table() function in R

To count the number of observations in each level of factor, we can use the R table() command as below:

```
> data(iris)
```

```
> head(iris)
```

```
> table(iris$Species)
```

```
> data(iris)
> head(iris)
  Sepal.Length Sepal.Width Petal.Length Petal.Width Species
1           5.1           3.5           1.4           0.2  setosa
2           4.9           3.0           1.4           0.2  setosa
3           4.7           3.2           1.3           0.2  setosa
4           4.6           3.1           1.5           0.2  setosa
5           5.0           3.6           1.4           0.2  setosa
6           5.4           3.9           1.7           0.4  setosa
> table(iris$Species)

  setosa versicolor virginica 
      50         50         50 
```

# merge() function in R

If you want to combine data from different sources in [R](#), you can combine different sets of data in three ways:

```
> data(cars)
> data(iris)
> str(cars)
'data.frame':  50 obs. of  2 variables:
 $ speed: num  4 4 7 7 8 9 10 10 11 ...
 $ dist : num  2 10 4 22 16 10 18 26 34 17 ...
> str(iris)
'data.frame':  150 obs. of  5 variables:
 $ Sepal.Length: num  5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
 $ Sepal.Width : num  3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
 $ Petal.Length: num  1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
 $ Petal.Width : num  0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
 $ Species      : Factor w/ 3 levels "setosa","versicolor",...: 1 1 1 1 1 1 1 1 1 1 ...
> merged <- merge(iris,cars)
> str(merged)
'data.frame':  7500 obs. of  7 variables:
 $ Sepal.Length: num  5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
 $ Sepal.Width : num  3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
 $ Petal.Length: num  1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
 $ Petal.Width : num  0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
 $ Species      : Factor w/ 3 levels "setosa","versicolor",...: 1 1 1 1 1 1 1 1 1 1 ...
 $ speed        : num  4 4 4 4 4 4 4 4 4 4 ...
 $ dist         : num  2 2 2 2 2 2 2 2 2 2 ...
> |
```

The *str()* command is designed to help you examine the structure of a data object rather than providing a statistical summary.

# Input/output

- **Input from keyboard**

```
> z <- scan()
```

- **You can use `readline()` for inputting a line from the keyboard in the form of a string:**

```
> w <- readline()  
bioinfo  
> w  
[1] "bioinfo"
```

- **In interactive mode, one can print the value of that variable by just typing the variable name or expression. In batch mode, one can use the `print()` function.**



# Input/output

- The function `read.table()` is used usually.
- The default value of a header is 'FALSE' and hence when you do not have a header, you need not say such.
- Basically, the character strings are considered as R factors. For turning this "feature" off, you can include the argument `as.is=T` in your call to `read.table()`.
- When you have a spreadsheet export file, i.e. having a type `.csv` where the fields are divided by commas in place of spaces, use `read.csv()` in place of `read.table()`.
- You can also use `read.xls` for reading core spreadsheet files.

# Exercise

- `a <- c(1,2,3,4,5,6,7,8)`
- `b <- c(3,4,5,6,7,8,10)`
- Calculate mean of a and b
- Check if `mean(a) > mean(b)` or not ?
- Download data from website
- Write a program to read data (`data_read3.txt`) and display content.

# summary() function in R

The *summary()* command will provide you with a statistical summary of your data.

```
> summary(iris)
```

```
> summary(iris)
  Sepal.Length   Sepal.Width   Petal.Length   Petal.Width   Species
Min.   :4.300   Min.   :2.000   Min.   :1.000   Min.   :0.100   setosa   :50
1st Qu.:5.100   1st Qu.:2.800   1st Qu.:1.600   1st Qu.:0.300   versicolor:50
Median :5.800   Median :3.000   Median :4.350   Median :1.300   virginica :50
Mean   :5.843   Mean   :3.057   Mean   :3.758   Mean   :1.199
3rd Qu.:6.400   3rd Qu.:3.300   3rd Qu.:5.100   3rd Qu.:1.800
Max.   :7.900   Max.   :4.400   Max.   :6.900   Max.   :2.500
> |
```

The `summary` command is, therefore, more useful as we can see minimum, maximum, mean, etc values. The `summary()` command works for both matrix and data frame objects by summarizing the columns rather than the rows.

# Name Commands in R

- Name command and its variants are used to find or add names to rows and columns of data structures.
- Below specified are few of the commands and their explanation:
  - `names()` – It works on matrix or data frame objects.
  - `rownames()` – It works on matrix or data frame objects and is used to give names to rows.
  - `colnames()` – It works on matrix or data frame objects and is used to give names to columns.
  - `dimnames()` – Gets row and column names for matrix or data frame objects, that is, it is used to see dimensions of the data frame.

# Exercise

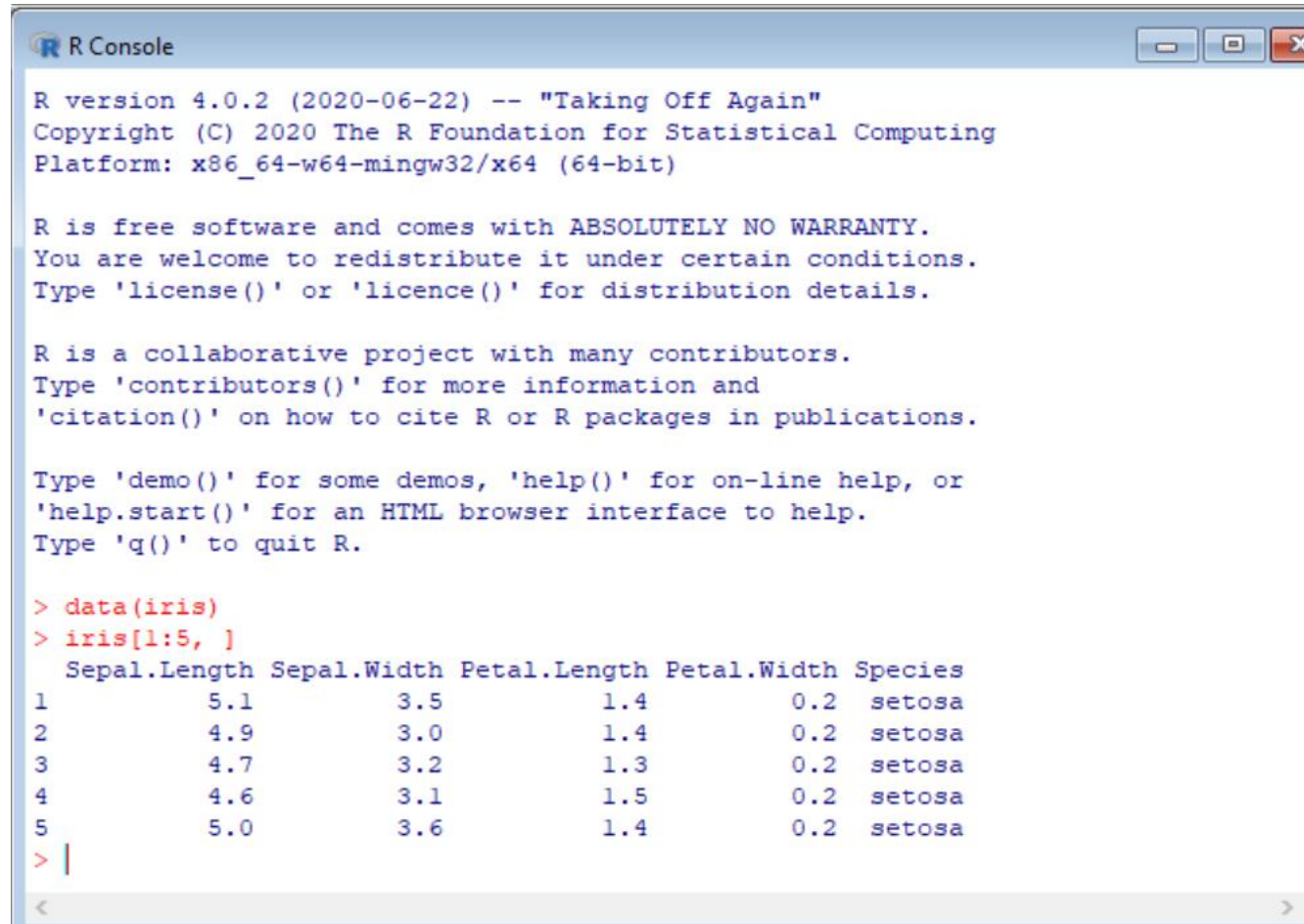
- Write a program to read data (data\_read3.txt) and display content.
- Display rownames() and colnames()

# Creating Subsets of Data in R

- The process of creating samples is called subsetting.
- As we know, data size is increasing exponentially.
- So, the data is divided into small-sized samples and analysis of samples is done.
- Different methods of subsetting in R are:
  - \$
  - The dollar sign operator selects a single element of data. The result of this operator is always a vector when we use it with a data-frame.
  - [[

- To retrieve 5 rows and all columns of already built-in dataset iris, the below command, is used:

```
> data(iris)
> iris[1:5, ]
```



```
R Console
R version 4.0.2 (2020-06-22) -- "Taking Off Again"
Copyright (C) 2020 The R Foundation for Statistical Computing
Platform: x86_64-w64-mingw32/x64 (64-bit)

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Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.

> data(iris)
> iris[1:5, ]
  Sepal.Length Sepal.Width Petal.Length Petal.Width Species
1           5.1           3.5           1.4           0.2  setosa
2           4.9           3.0           1.4           0.2  setosa
3           4.7           3.2           1.3           0.2  setosa
4           4.6           3.1           1.5           0.2  setosa
5           5.0           3.6           1.4           0.2  setosa
> |
```

# Data Visualization in R



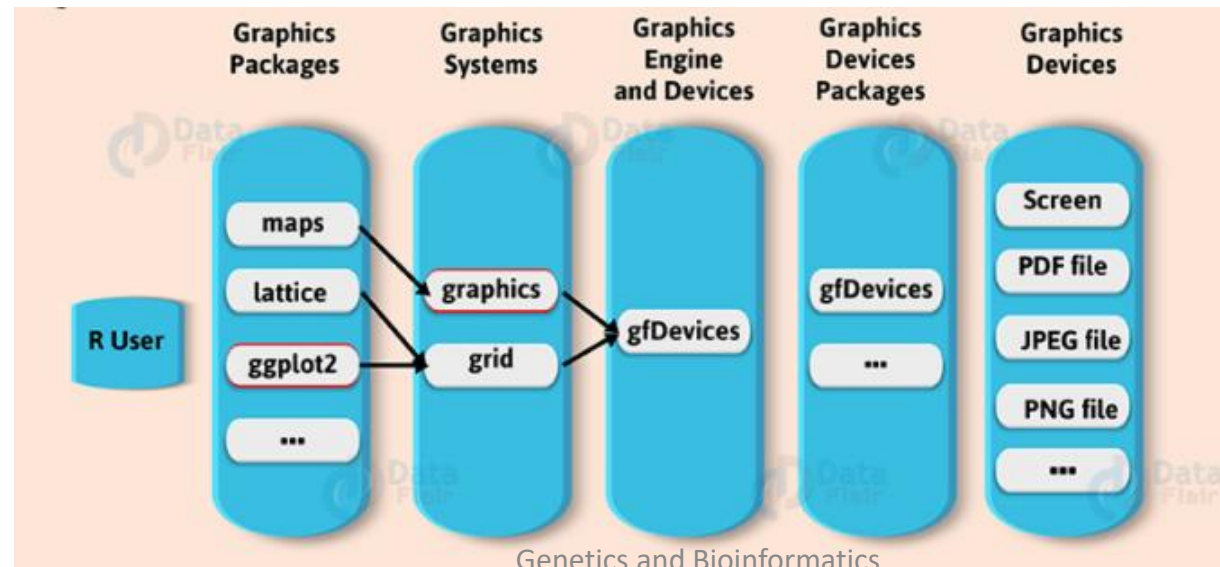
# Data Visualization in R

- **R Programming helps us to learn this art by offering a set of inbuilt functions and also libraries to build visualizations and present data.**
- **Before we move forward for the technical implementation of the visualization, let's see first how to select the right chart type.**

# Data Visualization in R - Simple

# Plotting in R

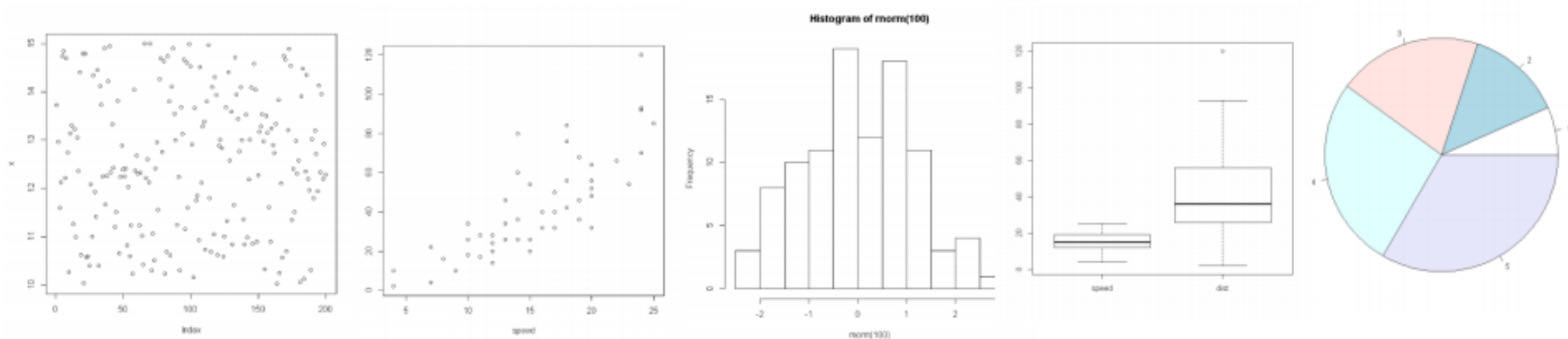
- R provides very rich set of plotting possibilities
- The basic command is `plot()`
- Each library has its own version of `plot()` function
- When R plots graphics , it opens ‘graphical device’ that could be either a window or file



# Plotting functions

- R standard graphics available through package graphics, include several functions that provide statistical plots, like:

Function	Description
<b>plot(x)</b>	plot of the values of x variable on the y axis
<b>plot(x,y)</b>	bi-variable plot of x and y values (both axis scaled based on values of x and y variables)
<b>pie(y)</b>	circular pie-char
<b>boxplot(x)</b>	Plots a box plot showing variables via their quantiles
<b>hist(x)</b>	Plots a histogram(bar plot)



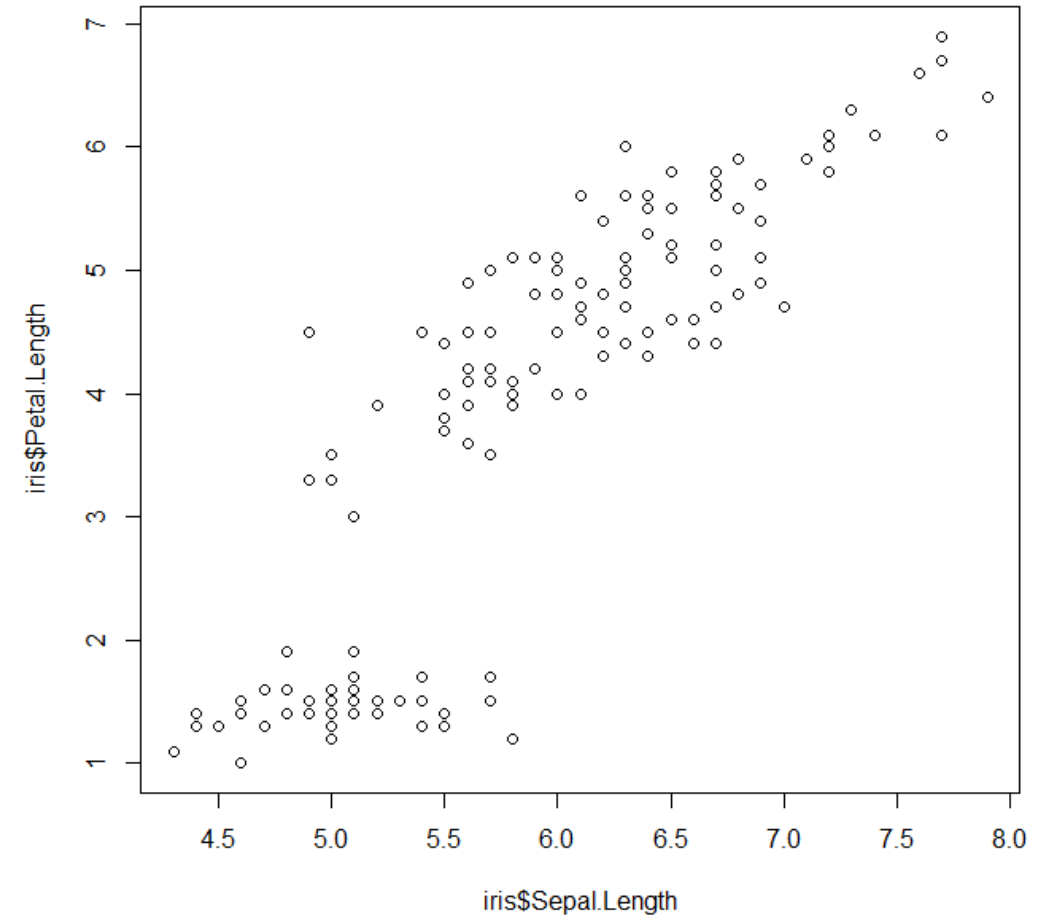
# Plot function

```
> data(iris)
```

```
> head(iris)
```

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
1	5.1	3.5	1.4	0.2	setosa
2	4.9	3.0	1.4	0.2	setosa
3	4.7	3.2	1.3	0.2	setosa
4	4.6	3.1	1.5	0.2	setosa
5	5.0	3.6	1.4	0.2	setosa
6	5.4	3.9	1.7	0.4	setosa

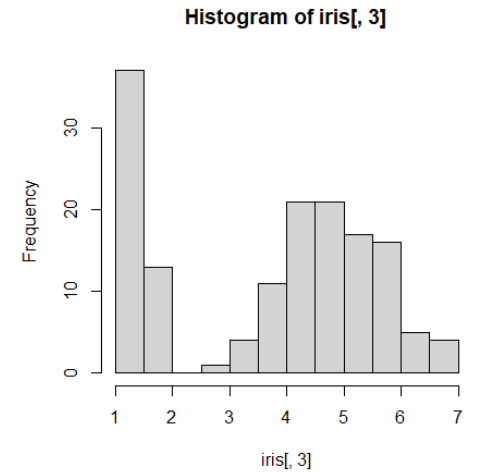
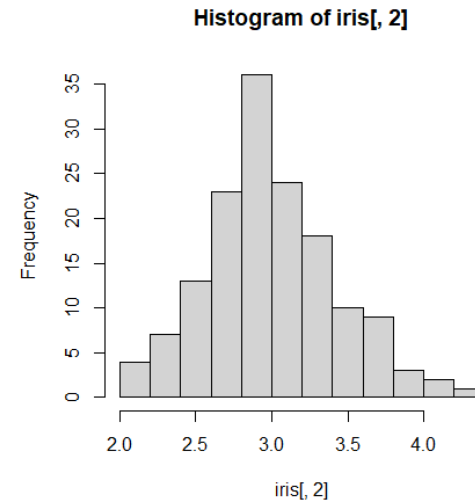
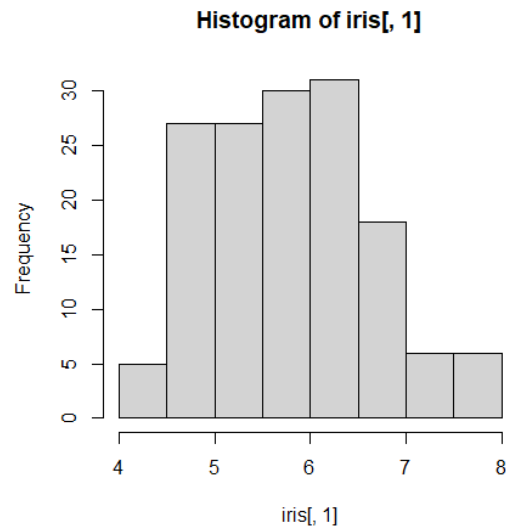
```
> plot(iris$Sepal.Length,iris$Petal.Length)
```



# Hist function

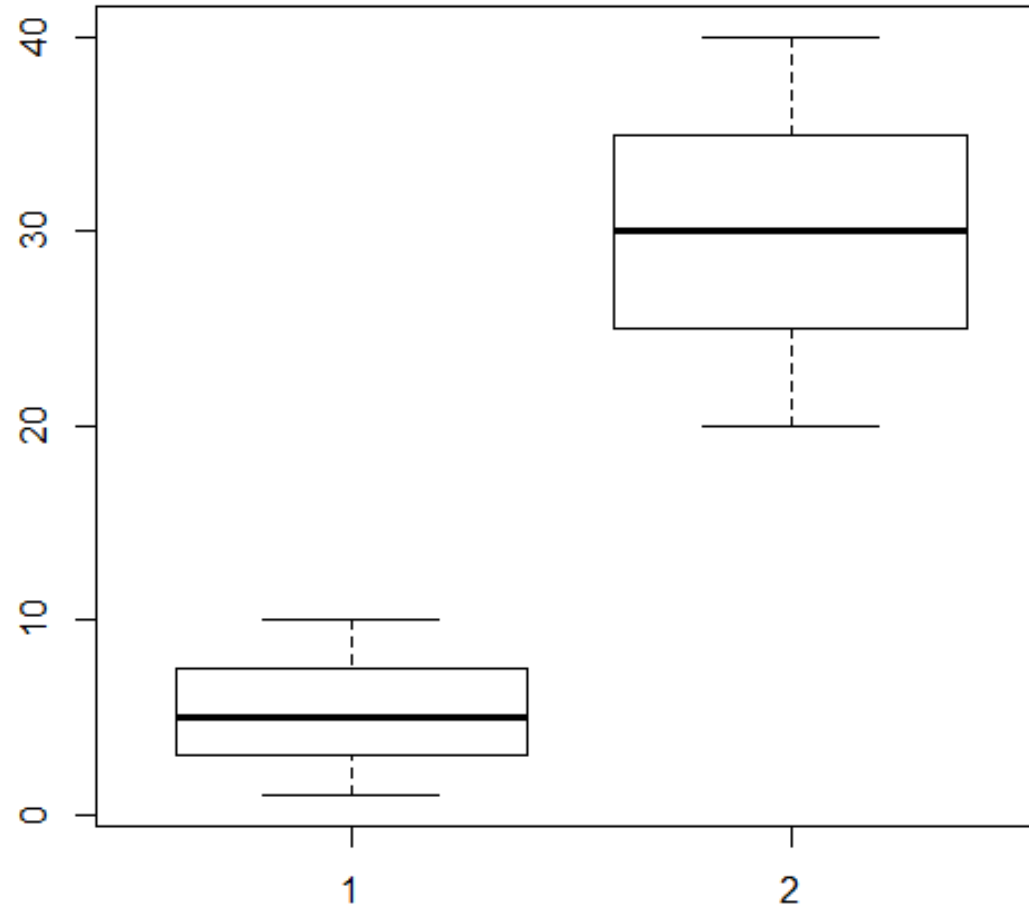
- Histogram
- A histogram is used to plot a continuous variable. Also, It helps to break the data into bins and shows the frequency distribution of these bins. Thus, we can always change the bin size and see the effect it has on visualization.

```
> hist(iris[,1])  
> hist(iris[,2])  
> hist(iris[,3])
```



# Boxplot function

```
> x <- c(1,5,10)  
> y <- c(20,30,40)  
> boxplot(x,y)
```

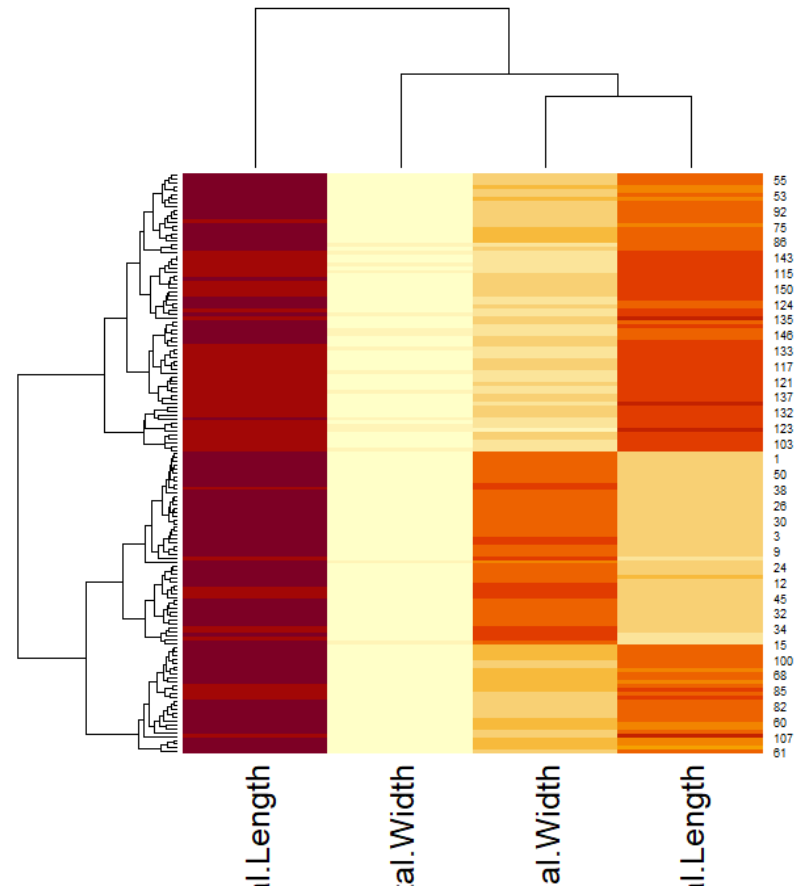


# Heatmap in R

- We use it for the intensity of colours. It is also used to display a relationship between two or three or many variables in a two-dimensional image. Thus, it allows us to explore two dimensions of the axis and the third dimension by an intensity of colour.

```
> iris_filtered <- iris[,1:4]
```

```
> heatmap(as.matrix(iris_filtered))
```



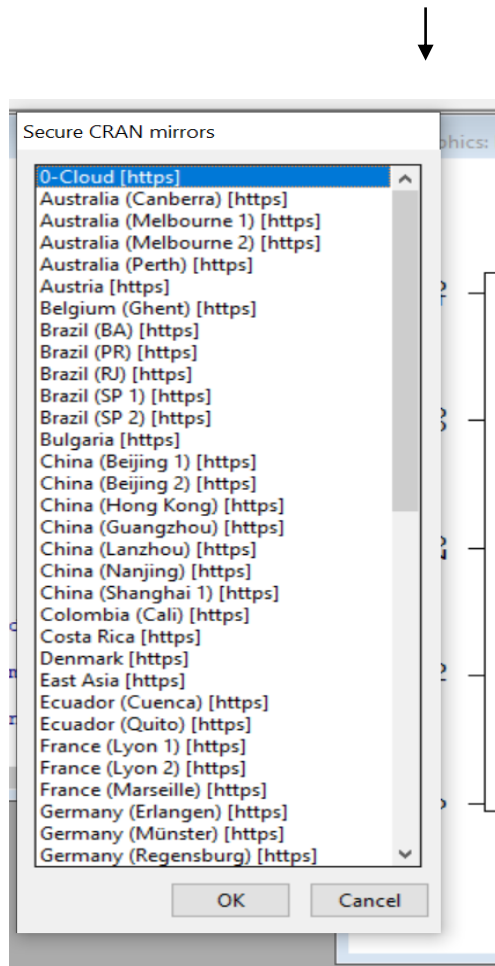


# Data Visualization in R - Advanced

# Library installation

**install.packages**

**> install.packages("ggplot2")**



```
> install.packages("ggplot2")
Installing package into 'C:/Users/archana/Documents/Administrator/R/win-library/3.6'
(as 'lib' is unspecified)
trying URL 'https://lib.ugent.be/CRAN/bin/windows/contrib/3.6/ggplot2_3.3.2.zip'
Content type 'application/zip' length 4068914 bytes (3.9 MB)
downloaded 3.9 MB

package 'ggplot2' successfully unpacked and MD5 sums checked

The downloaded binary packages are in
  C:\Users\archana\AppData\Local\Temp\Rtmpw5XZH3\downloaded_packages
```

**ggplot2 installed successfully!**

# Method to Save Graphs to Files in R

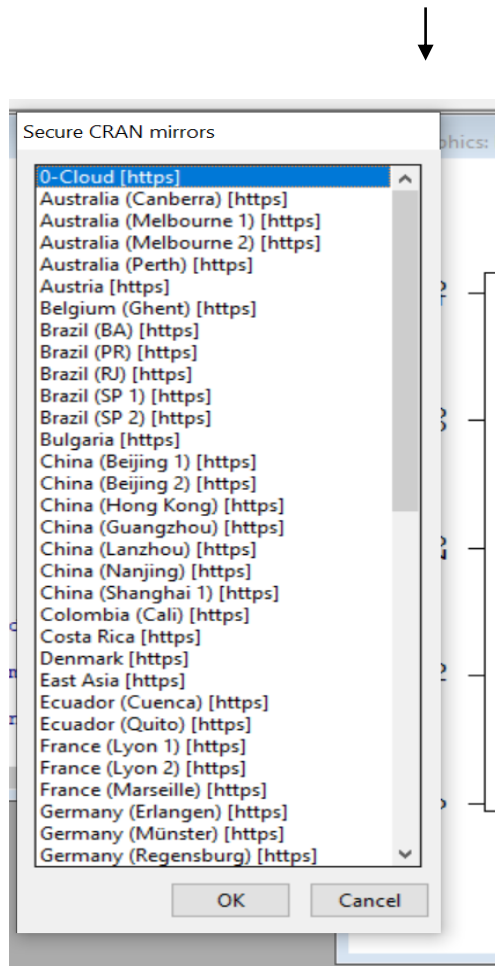
- In order to save graphics to an image file, there are three steps in R:
- You can create a graphics device of PNG format using `png()`, JPG format using `jpeg()` and PDF format using `pdf()`.
- Plot your data.
- Closing the graphics device and saving the image using `dev.off()`.

```
> jpeg("c:/mygraphs/myplot.jpg")  
  > x <- c(1,2,2)  
  > y <- c(3,4,4)  
  > plot(x,y)  
  > dev.off()
```

# Corrgram Library installation

**install.packages**

**> install.packages("corrgram")**



**Correlograms installed successfully!**

# Corrgram in R

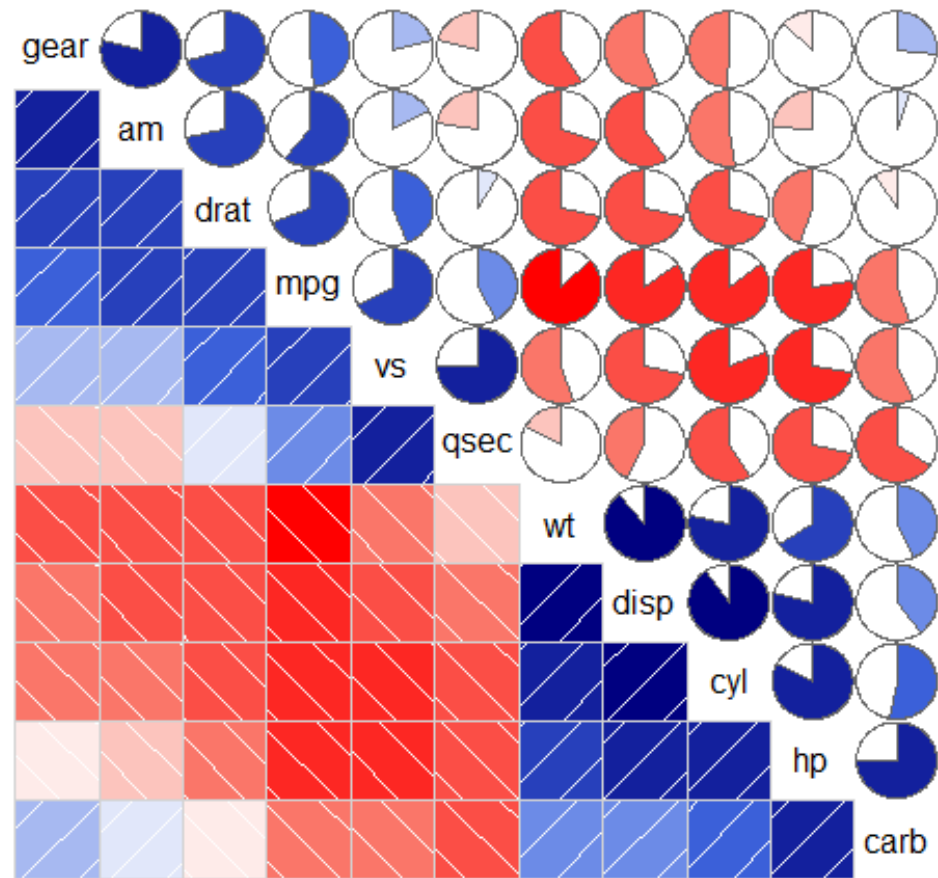
- We use it to test the level of correlation and also among the variable available in the dataset.
- Thus, the cells of the matrix can be shaded or coloured to show the co-relation value.

```
> library(corrgram)
```

```
> data(mtcars)
```

```
> corrgram(mtcars, order=TRUE, lower.panel=panel.shade, upper.panel=panel.pie,  
text.panel=panel.txt, main="Car Milage Data in PC2/PC1 Order")
```

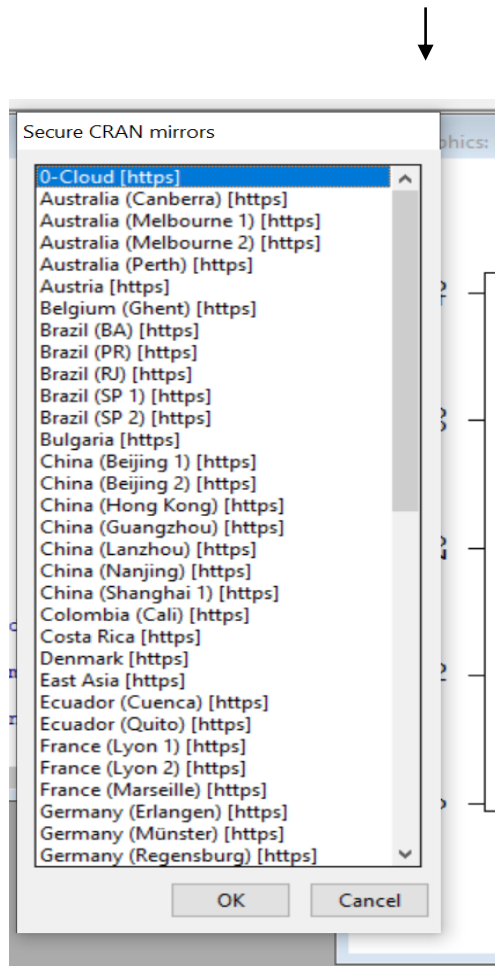
### Car Milage Data in PC2/PC1 Order



# circlize Library installation

**install.packages**

**> install.packages("circlize")**



**circlize installed successfully!**

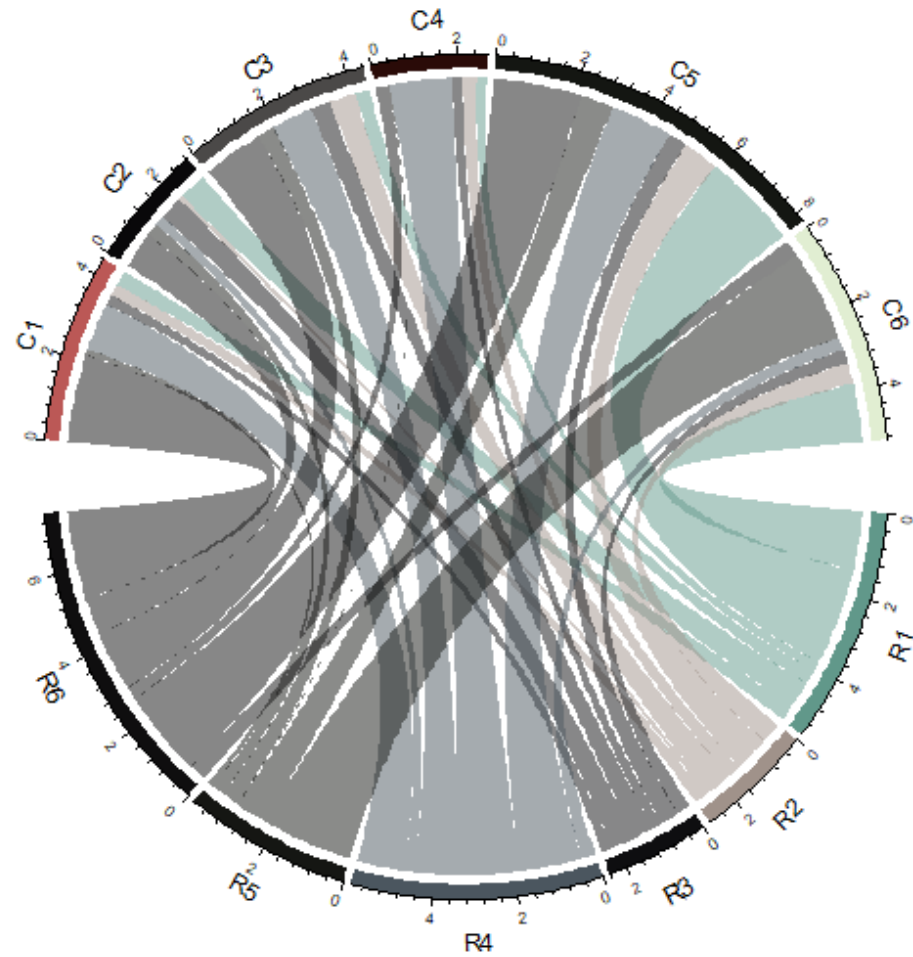
```
> library("circlize")
> mat = matrix(rnorm(36), 6, 6)
> rownames(mat) = paste0("R", 1:6)
> colnames(mat) = paste0("C", 1:6)
```

```
> mat = matrix(rnorm(36), 6, 6)
> rownames(mat) = paste0("R", 1:6)
> colnames(mat) = paste0("C", 1:6)
> mat
```

	C1	C2	C3	C4	C5	C6
R1	-0.3726024	0.6870889	0.3497039	0.24123038	-2.3779166	1.4447286
R2	0.2924820	0.1061324	-0.6347521	0.34156695	0.8794742	-0.5098149
R3	0.2520366	0.6003891	0.5031263	-0.25550267	0.4714030	0.3368118
R4	-1.1833159	-0.2451636	1.0311421	-1.55260575	1.5363097	0.3335470
R5	0.1313151	0.3054917	0.4118880	-0.02681157	0.8029791	2.2468392
R6	2.1757592	0.8985047	1.4857436	-0.30758183	-2.1677454	0.3984287

```
> chordDiagram(mat)
```



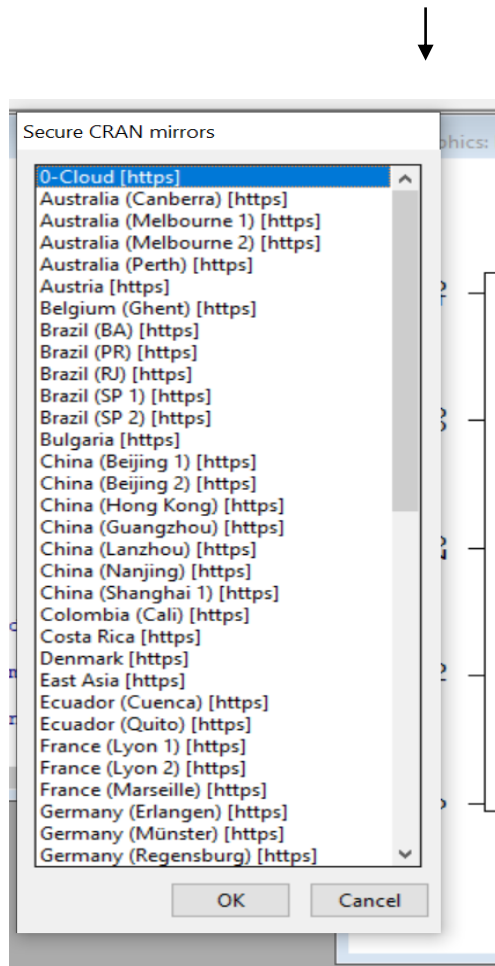


## matrix data in R

# igraph Library installation

**install.packages**

**> install.packages("igraph")**



**igraph installed successfully!**

## library(igraph)

*A graph is made up of vertices (also called nodes or points) which are connected by edges (also called links or lines).*

### #Define Nodes

```
nodes=cbind('id'=c('Fermenters','Methanogens','carbs','CO2','H2','other','CH4','H2O'),  
           'type'=c(rep('Microbe',2),rep('nonBio',6)))  
nodes
```

### #Define Links

```
links=cbind('from'=c('carbs',rep('Fermenters',3),rep('Methanogens',2),'CO2','H2'),  
           'to'=c('Fermenters','other','CO2','H2','CH4','H2O',rep('Methanogens',2)),  
           'type'=c('uptake',rep('output',5),rep('uptake',2)),  
           'weight'=rep(1,8))
```

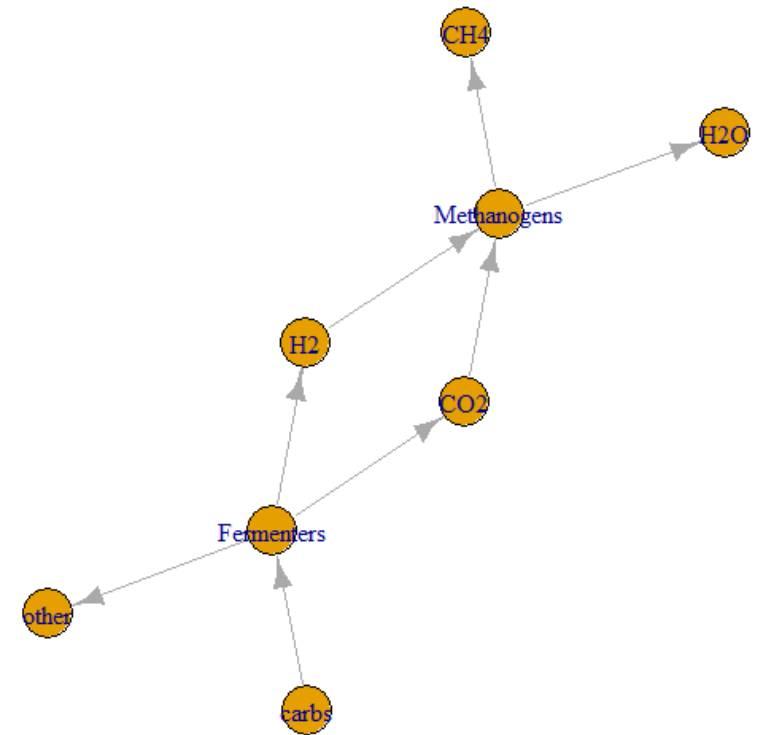
links

#Make the network

library(igraph)

net = graph\_from\_data\_frame(links,vertices = nodes,directed = T)

plot(net)



# Change Appearance

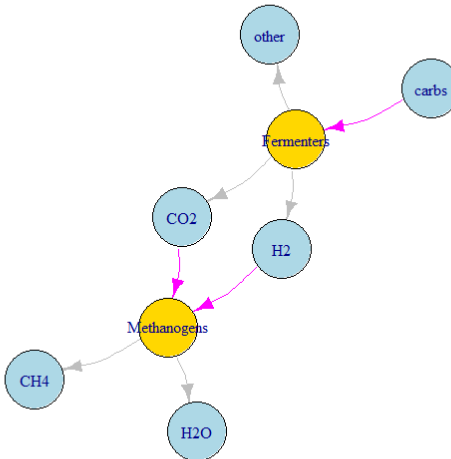
```
> colrs.v = c(nonBio = "lightblue", Microbe = "gold") #node colours
```

```
> V(net)$color = colrs.v[V(net)$type]
```

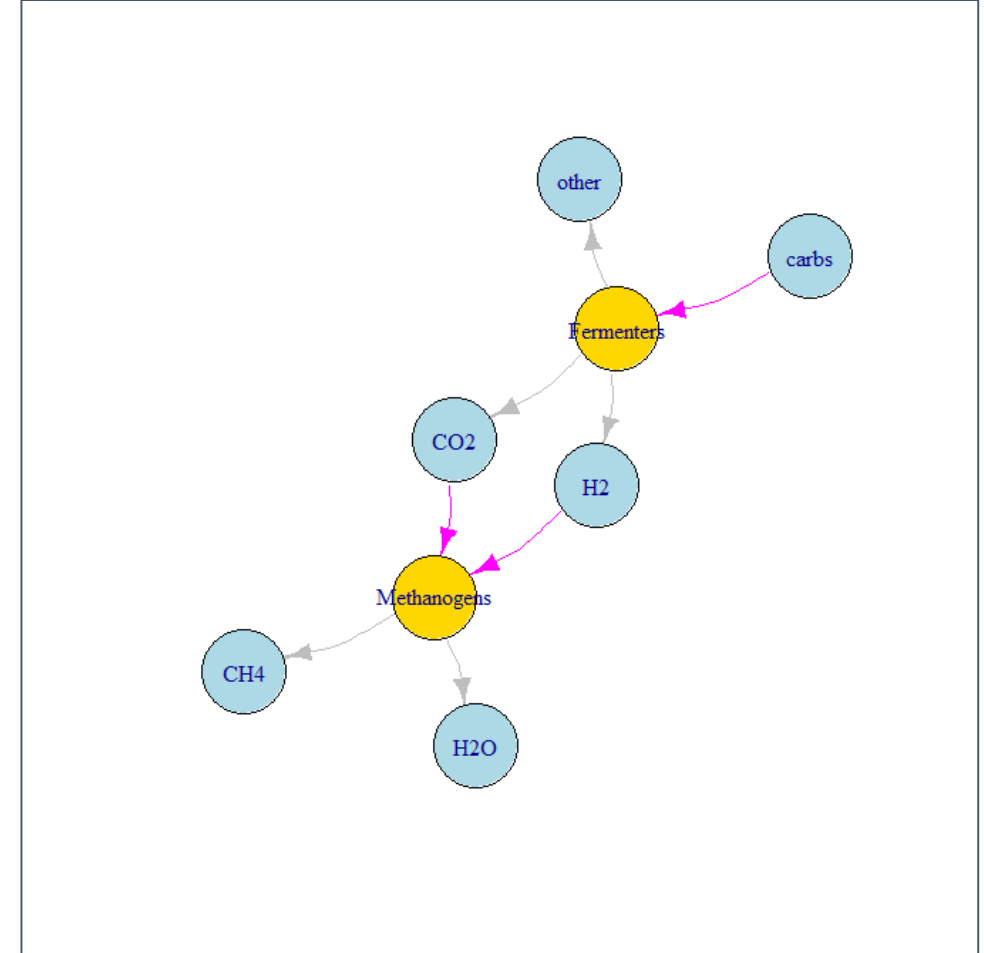
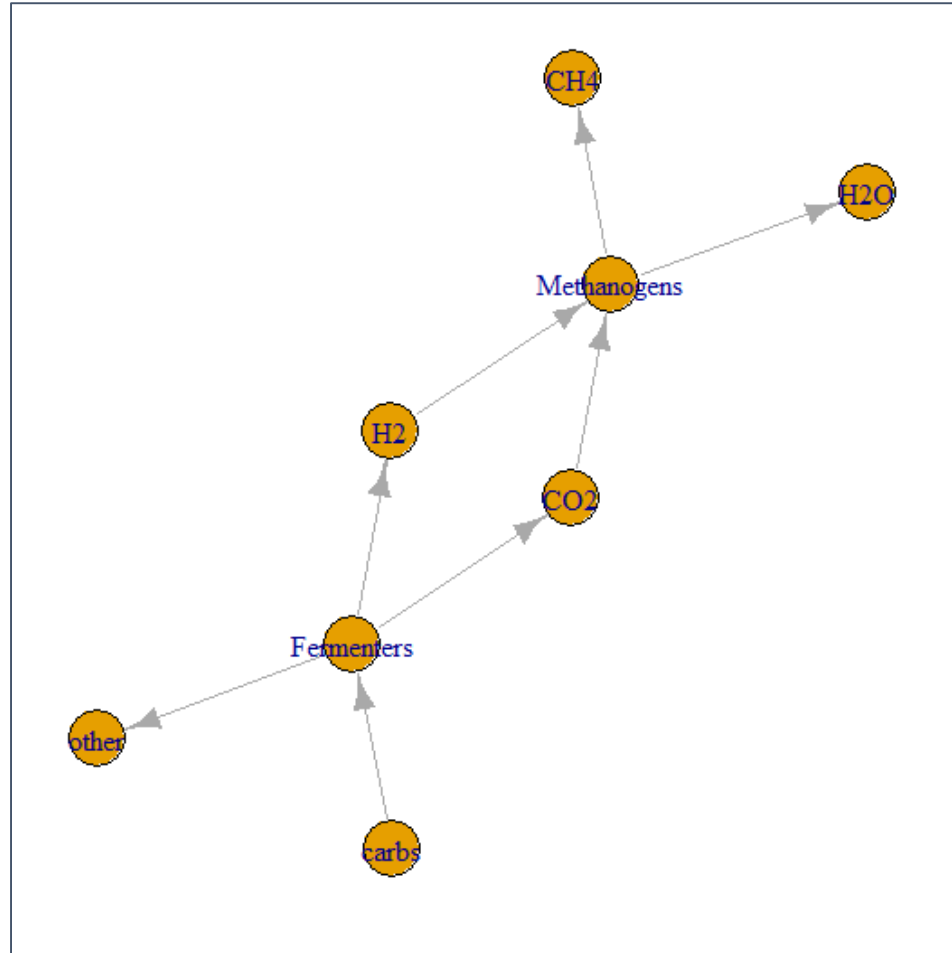
```
> colrs.e = c(output = "grey", uptake = "magenta") #edge colours
```

```
> E(net)$color = colrs.e[E(net)$type]
```

```
> plot(net, edge.curved=0.2, vertex.size=30) #make nodes bigger, curve arrows
```



# Comparison of both plots



# Density ridgeline plots

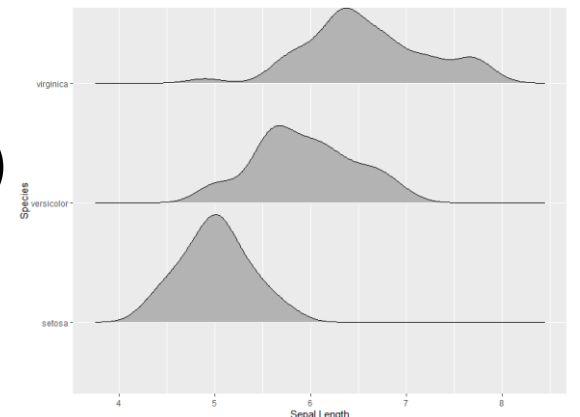
`library(ggridges)`

- The geom `geom_density_ridges` calculates density estimates from the provided data and then plots those, using the ridgeline visualization.

```
>ggplot(iris, aes(x = Sepal.Length, y = Species)) + geom_density_ridges()
```

- The extent to which the different densities overlap can be controlled with the `scale` parameter.
- A setting of `scale=1` means the tallest density curve just touches the baseline of the next higher one.
- Smaller values create a separation between the curves, and larger values create more overlap.

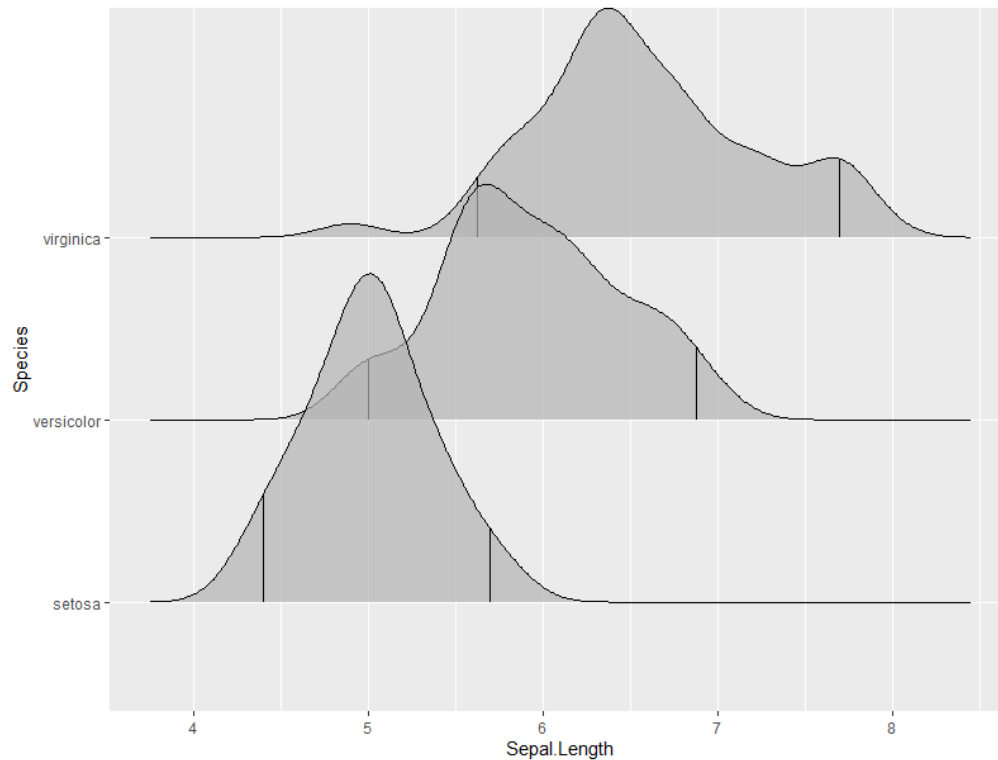
```
>ggplot(iris, aes(x = Sepal.Length, y = Species)) + geom_density_ridges(scale = 0.9)
```



# Density ridgeline plots

- We can also specify quantiles by cut points rather than number. E.g., we can indicate the 2.5% and 97.5% tails.

```
> ggplot(iris, aes(x = Sepal.Length, y = Species)) + stat_density_ridges(quantile_lines = TRUE, quantiles = c(0.025, 0.975), alpha = 0.7)
```

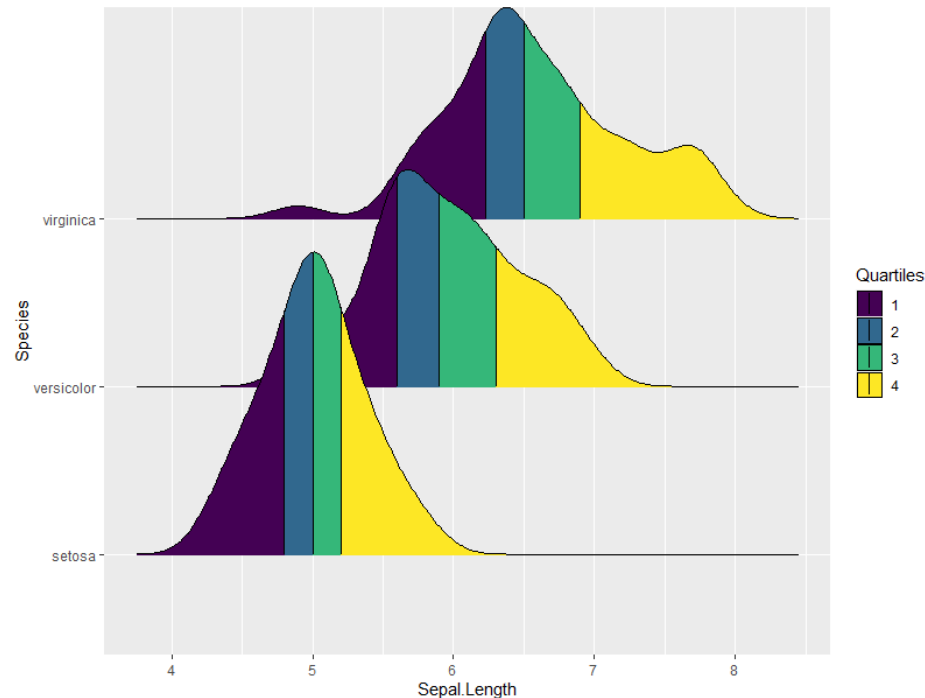




# Quantile plots

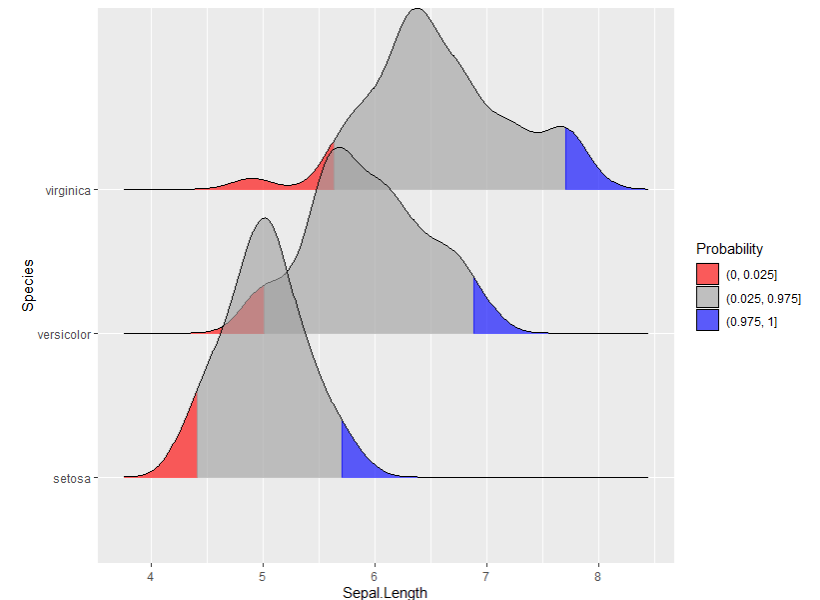
- Using the geom `geom_density_ridges_gradient` we can also color by quantile, via the calculated `stat(quantile)` aesthetic. Note that this aesthetic is only calculated if `calc_ecdf = TRUE`.

```
> ggplot(iris, aes(x=Sepal.Length, y=Species, fill = factor(stat(quantile)))) + stat_density_ridges( geom =  
"density_ridges_gradient", calc_ecdf = TRUE, quantiles = 4, quantile_lines = TRUE ) +  
scale_fill_viridis_d(name = "Quartiles")
```



- We can use the same approach to highlight the tails of the distributions.

```
> ggplot(iris, aes(x = Sepal.Length, y = Species, fill = factor(stat(quantile)))) +  
  stat_density_ridges(  
    geom = "density_ridges_gradient",  
    calc_ecdf = TRUE,  
    quantiles = c(0.025, 0.975)  
  ) +  
  scale_fill_manual(  
    name = "Probability", values = c("#FF0000A0", "#A0A0A0A0", "#0000FFA0"),  
    labels = c("(0, 0.025]", "(0.025, 0.975]", "(0.975, 1]")  
  )
```



# Data Transformation

# Dplyr package

- **dplyr is a grammar of data manipulation, providing a consistent set of verbs that help you solve the most common data manipulation challenges:**

- **The easiest way to get dplyr is to install the whole tidyverse:**

```
>install.packages("tidyverse")
```

- **Alternatively, install just dplyr:**

```
>install.packages("dplyr")
```

- **mutate() adds new variables that are functions of existing variables**
- **select() picks variables based on their names.**
- **filter() picks cases based on their values.**
- **summarise() reduces multiple values down to a single summary.**
- **arrange() changes the ordering of the rows.**

# Dplyr package

```
> head(starwars)
# A tibble: 6 x 14
  name      height  mass hair_color skin_color eye_color birth_year sex  gender
  <chr>    <int> <dbl> <chr>      <chr>      <chr>      <dbl> <chr> <chr>
1 Luke~    172    77 blond      fair        blue        19    male masculi~
2 C-3PO    167    75 <NA>      gold        yellow      112   none masculi~
3 R2-D2     96    32 <NA>      white, bl~ red         33    none masculi~
4 Dart~    202   136 none      white       yellow      41.9  male masculi~
5 Leia~    150    49 brown     light       brown       19    fema~ feminin~
6 Owen~    178   120 brown, gr~ light       blue        52    male masculi~
# ... with 5 more variables: homeworld <chr>, species <chr>, films <list>,
#   vehicles <list>, starships <list>
```

```
starwars %>% filter(species == "Droid")
```

- **Principal Component Analysis (PCA) is a useful technique for exploratory data analysis, allowing you to better visualize the variation present in a dataset with many variables.**
- **It is particularly helpful in the case of "wide" datasets, where you have many variables for each sample.**
- **Turning your original variables into a smaller number of "Principal Components"**

# Principal Component Analysis (PCA)

- **PCA is a type of linear transformation on a given data set that has values for a certain number of variables (coordinates) for a certain amount of spaces.**
- **This linear transformation fits this dataset to a new coordinate system in such a way that the most significant variance is found on the first coordinate.**
- **In this way, you transform a set of  $x$  correlated variables over  $y$  samples to a set of  $p$  uncorrelated principal components over the same samples.**
- **Where many variables correlate with one another, they will all contribute strongly to the same principal component.**

# Principal Component Analysis (PCA)

- **Let us use the mtcars dataset, which is built into R.**
- **This dataset consists of data on 32 models of car, taken from an American motoring magazine (1974 Motor Trend magazine). For each car, you have 11 features, expressed in varying units (US units).**
- **Compute the Principal Components**
- **Because PCA works best with numerical data, you'll exclude the two categorical variables (vs and am).**



# Principal Component Analysis (PCA)

- You are left with a matrix of 9 columns and 32 rows, which you pass to the `prcomp()` function, assigning your output to `mtcars.pca`.

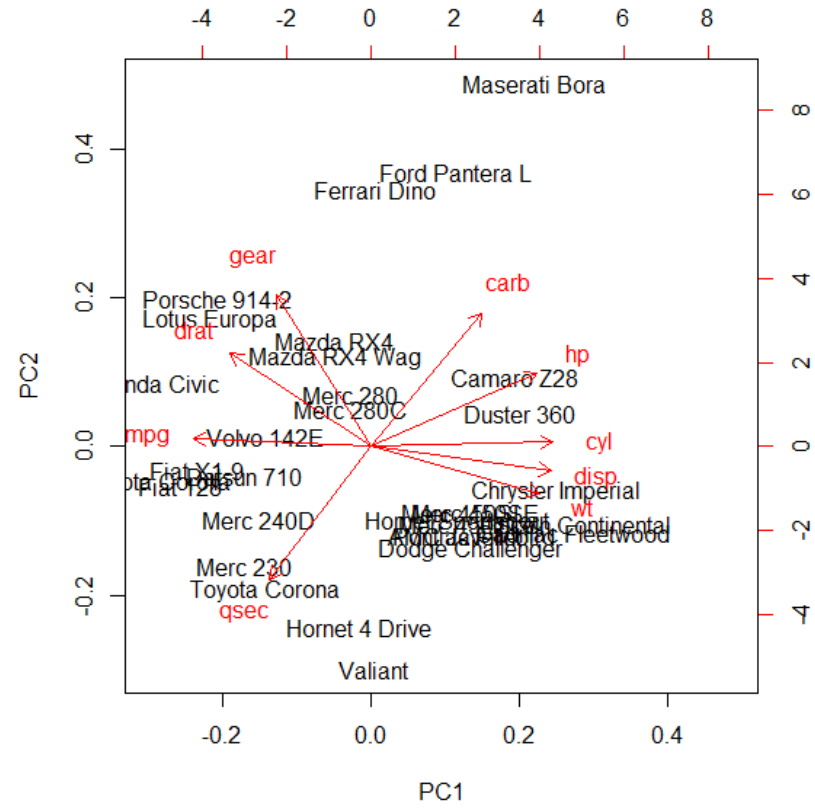
```
> mtcars.pca <- prcomp(mtcars[,c(1:7,10,11)], center = TRUE,scale. = TRUE)
```

- You will also set two arguments, `center` and `scale`, to be `TRUE`. Then you can have a peek at your PCA object with `summary()`.

```
> summary(mtcars.pca)  
>str(mtcars.pca)
```

- Let us draw biplot

`>biplot(mtcars.pca)`



# Exercise

- **install library ggfortify**
- **Load data iris**
  - **>data(iris)**
- **Perform the PCA analysis**
- **Develop biplot**
- **List highly correlated variables**
- **Display graph by use of following command :**

```
autoplot(iris.pca, data = iris, colour = 'Species')
```

```
autoplot(iris.pca, data = iris, colour = 'Species', label = TRUE, label.size = 3)
```

# R Linear Regression

- **What is Linear Regression?**

*Regression analysis is a statistical technique for determining the relationship between two or more than two variables.*

- **There are two types of variables in regression analysis –**

**independent variable and dependent variable.**

- **Linear Regression is of the following two types:**

- **Simple Linear Regression – Based on the value of the single explanatory variable, the value of the response variable changes.**

- **Multiple Linear Regression – The value is dependent upon more than one explanatory variables in case of multiple linear regression.**

# R Linear Regression

- Linear regression is one of the most basic statistical models out there.
- A linear regression can be calculated in R with the command `lm`.
- In the next example, use this command to calculate the height based on the age of the child.

$$\text{Height} = a + \text{Age} * b$$

- In this case, “a” and “b” are called the intercept and the slope respectively.
- With the same example, “a” or the intercept, is the value from where you start measuring.
- The slope measures the change of height with respect to the age in months. In general, for every month older the child is, his or her height will increase with “b”.

# Extracting information from a linear regression

- `x <- lm(y~x, data=z)`
- `summary(x)` comprehensive summary of results
- `print(x)` precise version of the object
- `deviance(x)` residuals
- `plot(x)` returns plots: residuals, fitted values and some diagnostics
- `coef(x)` extract regression coefficients

# R Linear Regression : example

Let us download the data to an object called `ageandheight` and then create the linear regression in the third line. The `lm` command takes the variables in the format:

*lm([target variable] ~ [predictor variables], data = [data source])*

```
> library(readxl)
```

```
> ageandheight <- read_excel("ageandheight.xls",  
sheet = "Hoja2")
```

```
> lmHeight = lm(height~age, data = ageandheight)
```

```
> summary(lmHeight) #Review the results
```

```
> ageandheight <- read_excel("ageandheight.xls", sheet = "Hoja2")  
> head(ageandheight)  
# A tibble: 6 x 3  
  age height no_siblings  
  <dbl> <dbl> <dbl>  
1  18  76.1         1  
2  19  77           2  
3  20  78.1         3  
4  21  78.2         2  
5  22  78.8         0  
6  23  79.7         1  
> lmHeight = lm(height~age, data = ageandheight)  
> summary(lmHeight)  
  
Call:  
lm(formula = height ~ age, data = ageandheight)  
  
Residuals:  
    Min       1Q   Median       3Q      Max   
-66.059   5.193   5.810   6.617   7.533   
  
Coefficients:  
            Estimate Std. Error t value Pr(>|t|)   
(Intercept)  64.8445    43.3815   1.495   0.166   
age           0.3835     1.8264   0.210   0.838   
  
Residual standard error: 21.84 on 10 degrees of freedom  
Multiple R-squared:  0.00439, Adjusted R-squared: -0.09517   
F-statistic: 0.0441 on 1 and 10 DF, p-value: 0.8379
```

```

Call:
lm(formula = height ~ age, data = ageandheight)

Residuals:
    Min       1Q   Median       3Q      Max
-66.059   5.193   5.810   6.617   7.533

Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept)  64.8445   43.3815   1.495   0.166
age           0.3835    1.8264   0.210   0.838

Residual standard error: 21.84 on 10 degrees of freedom
Multiple R-squared:  0.00439,    Adjusted R-squared:  -0.09517
F-statistic: 0.0441 on 1 and 10 DF,  p-value: 0.8379

```

## Coefficients.

In the orange square, you can see the values of the intercept (“a” value) and the slope (“b” value) for the age. These “a” and “b” values plot a line between all the points of the data.



# Exercise

- 1) Open Rstudio**
- 2) Run small code (given in exercise)**
- 3) Select option : html, pdf or doc**
- 4) Save file**
- 5) Show your reports**

# PLINK : GWAS DATA

# PLINK : Introduction

- **PLINK is a free, open-source designed to perform a range of basic, large-scale analyses in a computationally efficient manner.**
- **PLINK is whole genome association analysis tool.**
- **PLINK has a well documented manual.**
- **Available for linux, MAC and MAC-DOS.**
- **Command line version is faster than graphical PLINK.**

# PLINK : Multi-feature tool

- **Merge two or more files**
- **Extracts subsets (SNPs or individuals)**
- **Compress data in a binary file format**
- **PLINK has numerous useful features for managing and analyzing genetic data**
- **Read data in a variety of formats**
- **Recode and reorder files**

# Input Files

- **Genotype data is a text file**
- **Pedigree file (.ped)**
- **Map file (.map)**
- **Genotype data is a compressed binary file**
- **Fam File (.fam)**
- **Bim file (.bim)**
- **Bed file (.bed)**

# PED Input File

Pedigree File - the first six columns are mandatory:

- Family ID
- Individual ID
- Paternal ID
- Maternal ID
- Sex (1=male; 2=female; other=unknown)
- Phenotype

Column1	Column2	Column3	Column4	Column5	Column6				
1	1	0	0	1	1	A	A	G	T
2	1	0	0	1	1	A	C	T	G
3	1	0	0	1	1	C	C	G	G
4	1	0	0	1	2	A	C	T	T
5	1	0	0	1	2	C	C	G	T

# MAP Input File

MAP File has 4 columns:

- chromosome (1-22, X, Y or 0 if unplaced)
- rs# or snp identifier
- Genetic distance (morgans)
- Base-pair position (bp units)

Column1 Column2 Column3 Column4

1 snp1 0 1

1 snp2 0 2

# Others Input File

\*.ped

FID	IID	PID	MID	Sex	P	rs1	rs2	rs3
1	1	0	0	2	1	CT	AG	AA
2	2	0	0	1	0	CC	AA	AC
3	3	0	0	1	1	CC	AA	AC

\*.map

Chr	SNP	GD	BPP
1	rs1	0	870000
1	rs2	0	880000
1	rs3	0	890000

\*.fam

FID	IID	PID	MID	Sex	P
1	1	0	0	2	1
2	2	0	0	1	0
3	3	0	0	1	1

\*.bed

Contains binary version of the SNP info of the \*.ped file. (not in a format readable for humans)

\*.bim

Chr	SNP	GD	BPP	Allele 1	Allele 2
1	rs1	0	870000	C	T
1	rs2	0	880000	A	G
1	rs3	0	890000	A	C

Covariate file

FID	IID	C1	C2	C3
1	1	0.00812835	0.00606235	-0.000871105
2	2	-0.0600943	0.0318994	-0.0827743
3	3	-0.0431903	0.00133068	-0.000276131

Legend

FID	Family ID	rs{x}	Alleles per subject per SNP
IID	Individual ID	Chr	Chromosome
PID	Paternal ID	SNP	SNP name
MID	Maternal ID	GD	Genetic distance (morgans)
Sex	Sex of subject	BPP	Base-pair position (bp units)
P	Phenotype	C{x}	Covariates (e.g., Multidimensional Scaling (MDS) components)



- **Type command :**  
**plink --file test --noweb**

**Provide you detail information of SNPs count, individuals count and check for the missingness and frequency test**

```
Options in effect:
  --file test
  --noweb

2 (of 2) markers to be included from [ test.map ]
6 individuals read from [ test.ped ]
6 individuals with nonmissing phenotypes
Assuming a disease phenotype (1=unaff, 2=aff, 0=miss)
Missing phenotype value is also -9
3 cases, 3 controls and 0 missing
6 males, 0 females, and 0 of unspecified sex
Before frequency and genotyping pruning, there are 2 SNPs
6 founders and 0 non-founders found
Total genotyping rate in remaining individuals is 1
0 SNPs failed missingness test ( GENO > 1 )
0 SNPs failed frequency test ( MAF < 0 )
After frequency and genotyping pruning, there are 2 SNPs
After filtering, 3 cases, 3 controls and 0 missing
After filtering, 6 males, 0 females, and 0 of unspecified sex
```

# Data Transformation

- **Convert map and ped into binary format**

- *plink.exe --file test --make-bed --out test --noweb*

- **Read binary file in plink**

- *plink.exe -bfile test*

```
Reading map (extended format) from [ test.bim ]
2 markers to be included from [ test.bim ]
Reading pedigree information from [ test.fam ]
6 individuals read from [ test.fam ]
6 individuals with nonmissing phenotypes
Assuming a disease phenotype (1=unaff, 2=aff, 0=miss)
Missing phenotype value is also -9
3 cases, 3 controls and 0 missing
6 males, 0 females, and 0 of unspecified sex
Reading genotype bitfile from [ test.bed ]
Detected that binary PED file is v1.00 SNP-major mode
Before frequency and genotyping pruning, there are 2 SNPs
6 founders and 0 non-founders found
Total genotyping rate in remaining individuals is 1
0 SNPs failed missingness test ( GENO > 1 )
0 SNPs failed frequency test ( MAF < 0 )
After frequency and genotyping pruning, there are 2 SNPs
After filtering, 3 cases, 3 controls and 0 missing
After filtering, 6 males, 0 females, and 0 of unspecified sex

Analysis finished: Sun Oct 20 18:49:10 2019
```

# Comparison

```
2 (of 2) markers to be included from [ test.map ]
6 individuals read from [ test.ped ]
6 individuals with nonmissing phenotypes
Assuming a disease phenotype (1=unaff, 2=aff, 0=miss)
Missing phenotype value is also -9
3 cases, 3 controls and 0 missing
6 males, 0 females, and 0 of unspecified sex
Before frequency and genotyping pruning, there are 2 SNPs
6 founders and 0 non-founders found
Total genotyping rate in remaining individuals is 1
0 SNPs failed missingness test ( GENO > 1 )
0 SNPs failed frequency test ( MAF < 0 )
After frequency and genotyping pruning, there are 2 SNPs
After filtering, 3 cases, 3 controls and 0 missing
After filtering, 6 males, 0 females, and 0 of unspecified sex
```

**TEXT output**

```
Reading map (extended format) from [ test.bim ]
2 markers to be included from [ test.bim ]
Reading pedigree information from [ test.fam ]
6 individuals read from [ test.fam ]
6 individuals with nonmissing phenotypes
Assuming a disease phenotype (1=unaff, 2=aff, 0=miss)
Missing phenotype value is also -9
3 cases, 3 controls and 0 missing
6 males, 0 females, and 0 of unspecified sex
Reading genotype bitfile from [ test.bed ]
Detected that binary PED file is v1.00 SNP-major mode
Before frequency and genotyping pruning, there are 2 SNPs
6 founders and 0 non-founders found
Total genotyping rate in remaining individuals is 1
0 SNPs failed missingness test ( GENO > 1 )
0 SNPs failed frequency test ( MAF < 0 )
After frequency and genotyping pruning, there are 2 SNPs
After filtering, 3 cases, 3 controls and 0 missing
After filtering, 6 males, 0 females, and 0 of unspecified sex

Analysis finished: Sun Oct 20 18:49:10 2019
```

**BED OUTPUT**

# QC of genetic DATA

- **A vital step that should be part of any GWAS is the use of appropriate QC.**
- **Without extensive QC, GWAS will not generate reliable results because raw genotype data are inherently imperfect.**
- **Errors in the data can arise for numerous reasons, for example, due to poor quality of DNA samples, poor DNA hybridization to the array, poorly performing genotype probes, and sample mix-ups or contamination.**

# QC of genetic DATA

The seven QC steps consist of filtering out of SNPs and individuals based on the following:

- (1) individual and SNP missingness,**
- (2) inconsistencies in assigned and genetic sex of subjects (see sex discrepancy),**
- (3) minor allele frequency (MAF),**
- (4) deviations from Hardy–Weinberg equilibrium (HWE),**
- (5) heterozygosity rate,**
- (6) relatedness, and**
- (7) ethnic outliers (see population stratification).**

# Important Commands

Step	Command	Function
1: Missingness of SNPs and individuals	--geno	Excludes SNPs that are missing in a large proportion of the subjects. In this step, SNPs with low genotype calls are removed.
	--mind	<b>Excludes individuals who have high rates of genotype missingness. In this step, individual with low genotype calls are removed.</b>
2: Sex discrepancy	--check-sex	Checks for discrepancies between sex of the individuals recorded in the dataset and their sex based on X chromosome heterozygosity/homozygosity rates.
3: Minor allele frequency (MAF)	--maf	Includes only SNPs above the set MAF threshold.
4: Hardy–Weinberg equilibrium (HWE)	--hwe	Excludes markers which deviate from Hardy–Weinberg equilibrium.
5: Heterozygosity		Excludes individuals with high or low heterozygosity rates
6: Relatedness	--genome	Calculates identity by descent (IBD) of all sample pairs.
	--min	Sets threshold and creates a list of individuals with relatedness above the chosen threshold. Meaning that subjects who are related at, for example, $\pi\text{-hat} > 0.2$ (i.e., second degree relatives) can be detected.
7: Population stratification	--genome	Calculates identity by descent (IBD) of all sample pairs.
	--cluster --mds-plot k	<b>Produces a k-dimensional representation of any substructure in the data, based on IBS.</b>

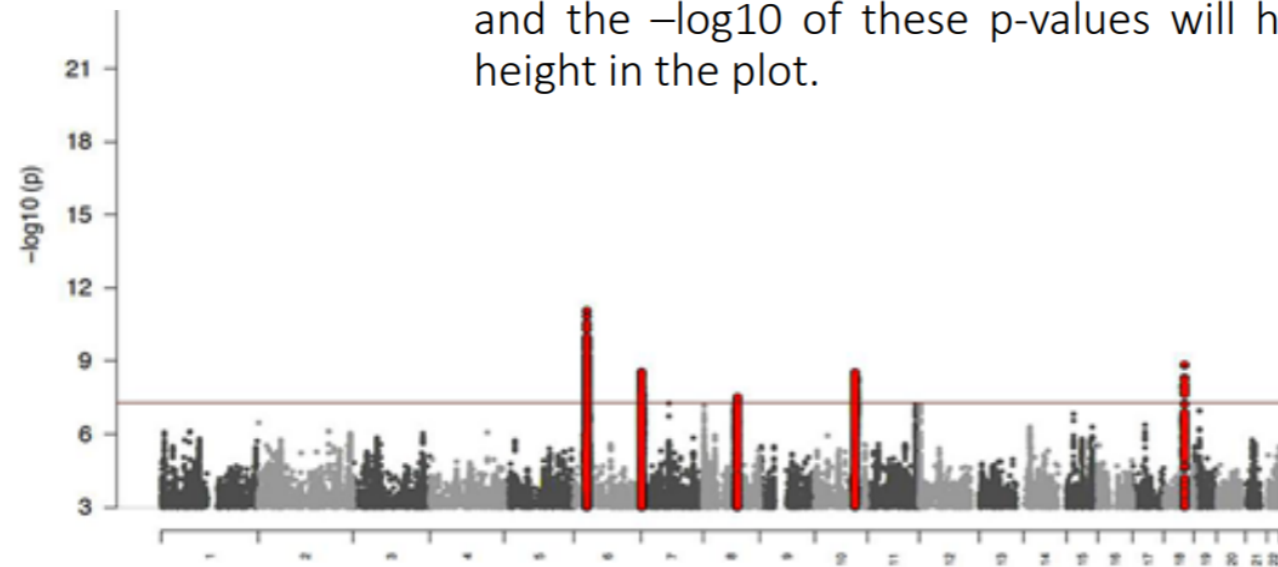
**-- assoc**

- **It works with case/control and continuous phenotypes.**
- **Case-control (1df chi-square test, outputs assoc)**
- **PLINK will recognise this is a case/control analysis because the phenotype just has:**
  - **1 (for controls),**
  - **2 (for cases), and**
  - **0/-9/non-numeric (for missing).**

- Given a quantitative phenotype, `--assoc` writes regression statistics and Wald test results to [plink.qassoc](#).

## Manhattan plot

- Plots the  $-\log_{10}$  of the association p-value for each SNP against the genomic coordinates.
- The strongest associations will have the smallest p-values and the  $-\log_{10}$  of these p-values will have the highest height in the plot.





# Useful Link

[https://jokergoo.github.io/circlize\\_book/book/](https://jokergoo.github.io/circlize_book/book/)

<https://igraph.org/r/doc/aaa-igraph-package.html>

<https://www.statmethods.net/advgraphs/correlograms.html>